

Review

Cell polarization, migration and tissue repair: A promising field for future melatonin research

Rüdiger Hardeland*

Johann Friedrich Blumenbach Institute of Zoology and Anthropology. University of Göttingen, Germany

*Correspondence: rhardel@gwdg.de

Running title: Polarization, migration and repair

Received: March 17, 2024; Accepted: July 2, 2024

ABSTRACT

Melatonin has been shown to support the repair of various tissue injuries. Wound healing is a complicated process that comprises several different cellular activities and regulation mechanisms, such as activation and programming of stem cells, interaction of different cell types, polarization of cells, especially concerning the alternative of pro- vs. anti-inflammatory behavior, migration of cells to the site of replacement, with guidance by other cells and modified extracellular matrix, as observed in the formation of biobridges. In most of these processes, melatonin acts as a decisive modulator, but details depend on tissue and cell types and have not been completely identified. Many aspects will require a considerable amount of work for understanding, in this context, the role of melatonin on a comprehensive basis. Moreover, the modulation of important cell properties has remained partially unknown or has only poorly considered in recent work. For instance, pro- or anti-inflammatory polarization of cells has been described in various cell types, not only in macrophages, in which melatonin is a major regulator, but also in microglia, in astrocytes and in neutrophils. Even in fibroblasts, polarization has been observed and concerns the alternative of inflammatory or fibrotic behavior. Notably, polarized cells that support healing in normal tissue seem to also protect tumors, whereas inflammatory phenotypes show antitumor activities. With regard to antitumor properties of melatonin, it seems necessary to clarify whether melatonin polarizes cells differently in the tumor microenvironment, compared to normal tissue, in which it promotes healing.

Key words: Biobridge, cell migration, cell polarization, healing, inflammation, melatonin, sirtuins, stem cells, tissue repair

1. INTRODUCTION

During the last decades, melatonin research had mainly focused on two important functions, melatonin's chronobiological role and the multiply documented protective actions of this fascinating molecule. The latter aspect mainly concerned its capacity of defending against free radicals, by both direct scavenging and reducing their formation via upregulating antioxidant enzymes. Additionally, melatonin was shown to be protective by contributing to the avoidance of physiological conditions that favor radical formation (1, 2). A more recent example for this latter capacity had emerged when melatonin was shown to modulate the polarization of

macrophages (3). This remarkable effect of favoring the polarization to anti-inflammatory M2 macrophages instead of the proinflammatory M1 phenotype resolved the previous paradox that melatonin can act in either pro- or anti-inflammatory ways, depending on conditions (4). The changes in polarization can be easily followed by determining the M1 marker iNOS (inducible NO synthase) and the M2 marker Arg-1 (arginase 1) (3). As far as melatonin acts on macrophages that had been polarized before to the M1 type or on stable cell lines derived from them, it will act in a proinflammatory way (5). However, this will not take place, if the cells had previously been polarized to M2. This kind of functional differentiation towards M2 cells may explain why melatonin was frequently shown to act *in vivo* as an anti-inflammatory agent (4, 6-8). However, the development of M2 macrophages bears an additional complexity, as several subtypes have to be distinguished, which differ with regard to their functional context. The subtype M2a is typically found to be concomitantly active with M1 and, therefore, seems to serve as a preventing agent against overshooting M1 activity. The other subtypes are associated with wound healing, in which M2b is typical for the proliferative phase after injury, whereas M2c appears in a later phase of remodeling or regeneration (9). An additional subtype, M2d, was found to be induced by the typically but not generally proinflammatory IL-6 via STAT3 (10). Moreover, the abundance of subtypes is, to a certain extent, tissue-specific and variable with regard to diseases (11, 12). It should also be noted that, in later stages of wound healing, melatonin may also increase iNOS again, in conjunction with an upregulation of VEGF (vascular endothelial growth factor) (13). The increase in iNOS may not imply a reversal of M2 polarization, but rather a replacement of macrophage subpopulations. The rise in VEGF can be interpreted as an angiogenic stimulus. Importantly, the different M2 subtypes have not generally to be seen as the friendly factors as they may appear because of their anti-inflammatory properties. In the context of cancer biology, they have turned out to be tumor promoting, contrary to the anti-tumor activities of M1 macrophages (12, 14). With regard to melatonin, which also acts differently in tumor and nontumor cells (15), though in an opposite way, it might be of interest to find out whether it may not favor M2 polarization in cancer cells.

Regardless of these details, the association of M2 macrophages with healing indicates another possible role of melatonin in processes of tissue repair. In fact, melatonin's potential in tissue repair has been observed and discussed in various organs (16-18), either in terms of direct effects, or via modulation of other signaling molecules, or being mediated by exosomes from melatonin-treated cells (17). Not surprisingly, these observations also included the participation of stem cells that are needed for the repair processes. The modulation of stem cell programming by melatonin has been recently reviewed (19).

Another important aspect of cell polarization, again with consequences to tissue repair, can be found in the central nervous system, insofar as this phenomenon also exists in microglia, in a similar way as in macrophages, and may exceed to other cell types that are usually not discussed in the context of polarization (20).

Tissue repair frequently comprises the necessity of cell migration in order to place the newly differentiated cells to the sites of requirement. This aspect, which is tightly associated with differential stem cell programming and also influenced by polarization of regulator cells, is of particular interest for repair in the central nervous system (19). Initial findings indicate that melatonin participates in these intertwined mechanisms.

This short review does not primarily focus on the contributions of melatonin released from numerous different cell types including immune cells (2, 21) and interactions between its different sources (22-24), but rather on aspects of applicability, either by administration of melatonin or exosomes from melatonin-treated macrophages.

2. PROGRAMMING OF STEM CELLS

Melatonin has been repeatedly shown to participate in the programming of various types of stem cells, as recently summarized (19). Comparisons of the different types of stem cells lead to the necessity of distinguishing between specific modes of action, not only with regard to the varying differentiation factors involved. In particular, the spectrum of effects differs profoundly between unipotent and pluripotent stem cells. Pluripotent stem cells can be selectively driven by external regulators such as melatonin towards specific developmental directions, as has been shown in the classic case of mesenchymal stem cells (MSCs) that can switch between adipogenic, chondrogenic and osteogenic routes. Depending on the cellular environment, high or low tissue-specific expression of transcription factors such as Pax2 (paired box 2) and Egr1 (early growth response protein 1), of differentiation factors such as Wnt (a name derived from the *Drosophila* gene *wingless* and the murine virus integration gene *Int-1*) and its downstream mediators, especially β -catenin, melatonin can promote any of these routes, but, in the presence of these regulators, it typically favors osteogenicity (19, 25-30). Additionally, differential epigenetic modifications of CpG islands in *Runx2* and *PPAR γ* promoters seem to be relevant to the allowance of the respective alternate route (31).

However, it is important to be aware that these three developmental routes of the MSC fate mainly represent examples devoid of exclusiveness. These findings are mainly based on experiments using adipose tissue-derived and bone marrow MSCs. MSCs from other sources, such as inflamed dental pulp or umbilical cord, and even myoblasts can also be directed towards osteogenesis, if Wnt4 or other Wnt subforms are upregulated independently of melatonin (19). On the other hand, MSCs from adipose tissue, bone marrow, amniotic fluid and dental pulp can be transdifferentiated to neuronal precursor cells in an alternate ectopic environment, presumably based on noncanonical modes of Wnt signaling (19).

Contrary to the potential of multilateral differentiation in pluripotent stem cells, unipotent stem cells can only develop in a unilateral mode. Nevertheless, regulator molecules such as melatonin can have a substantial impact on the fate of these cells. Melatonin has been shown to act in multiple ways on such cells, as observed in spermatogonial stem cells. Apart from protecting against chemical insults caused, e.g., by oxidative stress, enhanced mitochondrial activity and its preservation have been recently shown to be decisive for stem cell proliferation, which is impaired under conditions of mitochondrial stress (32). With regard to melatonin, two aspects should be, in this context, of particular interest. First, the multiply demonstrated protective actions of melatonin on mitochondria (8, 33-39) might facilitate or contribute to stem cell activation. Second, melatonin was shown to upregulate, in nontumor cells, several sirtuins, especially SIRT1 and SIRT3 (40-42), while functionality of these sirtuins has been found to be a prerequisite for the avoidance of mitochondrial stress (32). Moreover, melatonin can promote differentiation of spermatogonial stem cells to sperm-like cells and enhance their proliferation rates *in vivo*, in particular, via stimulation of GDNF (glial cell line-derived neurotrophic factor) release by Sertoli cells (43). As far as asymmetrical cell division of stem cells is stimulated, melatonin also supports the self-renewal of the stem cell population and, thereby, antagonizes their age-dependent declines. Moreover, self-renewal is typically associated with the conservation of stemness, as usually demonstrated by nestin expression levels (19, 44, 45). Such effects have been observed in both unipotent and pluripotent stem cells and are, therefore, independent of programming by melatonin (19).

3. POLARIZATION OF CELLS

As already mentioned in the introduction, polarization to cell subtypes with partially or completely different properties is not restricted to macrophages, but also occurs in microglia

(19). However, similar phenomena exist in other cells (Figure 1), but are only rarely considered. For instance, astrocytes can exist as two functionally different subtypes, known as neurotoxic A1 and neuroprotective A2 cells (46-48). Like M1 microglia, A1 astrocytes are believed to release NO, TNF α , IL-6, IL-1 β , the same CCL and CXCL chemokines, and additionally IL-1 α , GM-CSF and the complement protein C1q, whereas A2 cells secrete the anti-inflammatory cytokines IL-4, IL-10, IL-13 and TGF β (46). The signals driving to A1 or A2 are partially related to microglial activities, whereas anti-inflammatory polarization to A2 can be also achieved via ER (estrogen receptor) signaling, otherwise known for its neuroprotective effects (49). According to actual knowledge, it seems that polarizations to proinflammatory M1 and A1 or, alternately, to the anti-inflammatory M2 and A2 cells are jointly regulated and mechanistically interconnected, depending on local requirements. This may be favorable, as the same tissue or parts of it can be influenced towards the same direction. A presumably important aspect of the interconnections concerns the polarizing effects of cytokines that are released from other already polarized cells. For instance, TGF β released by M2 macrophages or M2 microglia favors polarization of other cell types to the anti-inflammatory phenotype. Similar effects may be assumed for other cytokines. Insofar, melatonin can be expected to mediate polarization by additional indirect effects, as a direct influence that leads to M2 macrophages or M2 microglia should be able to induce anti-inflammatory phenotypes in the other cell types, too (cf. Figure 1).

The existence of pro- and anti-inflammatory cell types that are generated by polarization has been also described in neutrophils, which primarily act in the periphery. Correspondingly, they are named N1 and N2 (50). The properties of the two cell types seem to partially differ with regard to organs and microenvironment. In the oral cavity, their polarization seems to be influenced by the local microbiota (50). As indicated above, polarization to anti-inflammatory N2 cells is favored by TGF β (51, 52), whereas the spectrum of N1 inducing cytokines awaits further investigation. With regard to cancer biology, N1 and N2 exhibit properties reminiscent of M1 and M2 macrophages (cf. Introduction). Tumor-associated neutrophils (TANs) polarized to the proinflammatory N1 type (N1 TANs) are anti-tumorigenic, in particular, by expressing immune-activating cytokines and chemokines, whereas anti-inflammatory N2 TANs have proved to be protumorigenic by suppressing tumor-directed immune responses and releasing angiogenic factors (50). The spectrum of neutrophil subtypes seems to exceed the N1/N2 polarity. Subsets that are preferentially recruited to sites of inflammation may be partially, but not completely identical with N1 cells.

A typical difference between cells that are polarized to either a proinflammatory or an anti-inflammatory phenotype concerns the expression of either inducible NO synthase (iNOS) or arginase 1 (Arg1) (3), observations that are not only applicable to monocyte-derived cell types. In addition to its involvement in oxidative and nitrosative stress, NO acts as an inflammatory signaling molecule (1-4, 7). Therefore, NO release by M1, A1, and N1 cells is the consequence of iNOS activation, whereas its lack in M2, A2 and N2 results from the consumption of the NOS substrate, arginine, by Arg1. Thereby, both oxidative/nitrosative stress and inflammatory signaling are strongly reduced. Moreover, arginine degradation is the starting point of polyamine synthesis, i.e., formation of another group of anti-inflammatory molecules (53).

Generally, polarization of cells to generate functionally different subsets should not be regarded as an exception, but seems to be a rather common phenomenon. This may even concern cells that are, from a conventional point of view, not considered as major players in inflammation control and redox biology. Fibroblasts may serve as an example. These cells are mostly perceived as structural components of tissues, including the stroma of tumors.

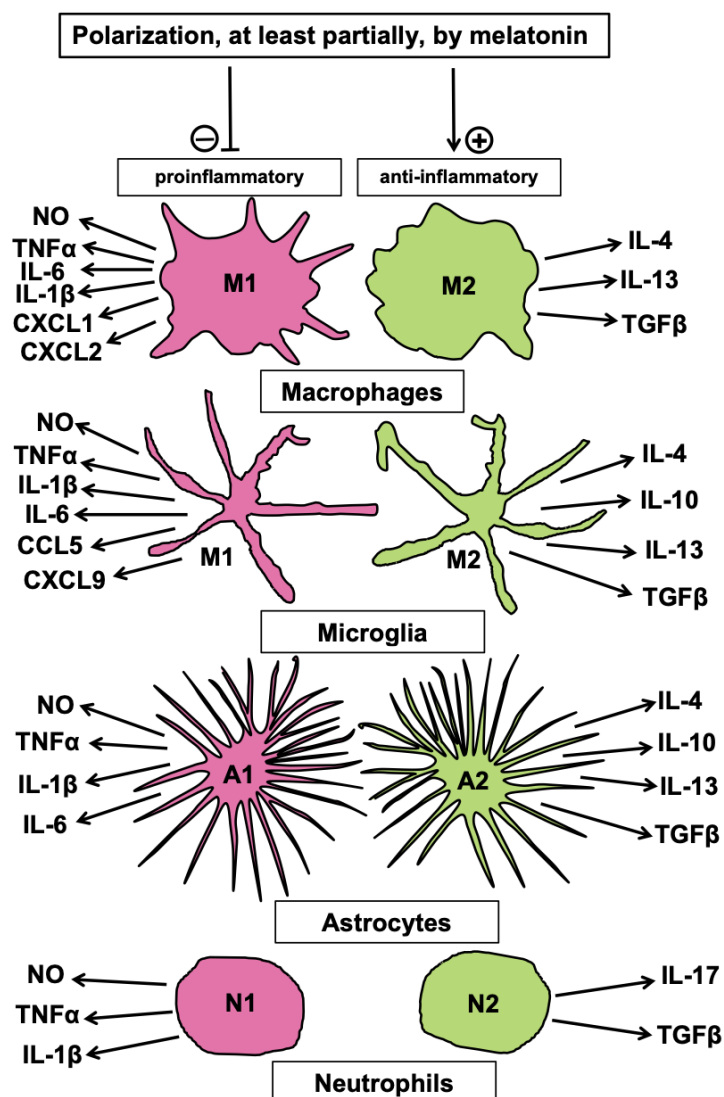


Fig. 1. Proinflammatory vs. anti-inflammatory polarization in four different cell types.

IL, interleukin; NO, nitric oxide; TGF β , transforming growth factor β ; TNF α , tumor necrosis factor α ; CCL, CXCL, chemokines. The numbers of cytokines and chemokines released from the respective polarized phenotypes may be even larger, but this would require further investigation or confirmation.

The tissue-specific microenvironment can exert different influences on fibroblasts by exposing them to locally specific factors released by neighboring cells or to components of the extracellular matrix. Such differences between induced subtypes become particularly evident when comparing tumor and nontumor tissues. Changes in stromal properties between tumor suppression and tumor promotion can be observed, in particular, in the fibroblasts, which represent a substantial quantity of the tumor stroma. Under the influence of cancer-derived cytokines, normal fibroblasts turn to cancer-associated fibroblasts (CAFs). Under these conditions, an immunosuppressive microenvironment is generated that favors tumor growth and has been reported to be explainable by the expression of the tumor-promoting gene LOXL2 (54). It was also reported that the expression of HIF-2 in CAFs contributes to the immunosuppressive microenvironment by polarizing macrophages to M2 in pancreatic cancer (55). Again, this conclusion underlines the importance of interplays between polarized cells of

different origin. In normal tissue, fibroblasts can be polarized to two main phenotypes, an inflammatory and a fibrotic one. In mouse fibroblasts, this was achieved by exposure to TNF α or TGF β 1, respectively (56). As either of these directions of polarity can be problematic in pathological terms, the two phenotypes should not be regarded as a usual alternative in healthy tissues, but rather as possible consequences of fibroblast activation.

Presumably, the list of polarizable cells will become much longer upon progressing research in this field. However, polarization does not necessarily concern the alternative of pro- vs. anti-inflammatory properties. In a more general understanding, any induced, at least moderately stable functional diversification of cells may be considered as a result of a kind of polarization, Especially the immune system, with its countless cellular subspecies and cytokine-induced phenotypes, will reveal many cases in which polarization, in the broader sense, takes place.

4. CELL MIGRATION

Stem cell activation and polarization of differentiating cells are frequently associated with migration. In particular, this is a requirement for the repair of injured tissues, since cells need to be directed to the site of damage for purposes of replacement. Cell migration can be observed in different cells during repair of various tissues. In the skin, keratinocytes, hair follicle cells and fibroblasts can be shown to migrate (57-59). Migratory fibroblasts are discernable from their resting stages by changes in the intracellular position of the nucleus (60). With regard to the involvement of SIRT1 in these cases (59, 60), effects via melatonin may be assumed. However, even under ectopic conditions, melatonin stimulated migration of endothelial stem cells in a study on renal ischemia, in addition to maturation effects (61). A recent review addressed the relationships between differentiation, proliferation and migration of cells after a preceding inflammatory phase with autophagy, followed by new formation of extracellular matrix and macrophage polarization to M2 (62). According to this view, inflammation and autophagy are required for local pathogen clearance, but also provide signals for activation and migration of cells as well as healing processes including M2 polarization. Another study interpreted the stimulation of migration in adipose-derived stem cells by melatonin as a consequence of downregulation of proinflammatory cytokines (63).

An important aspect of cell migration is the role of the extracellular matrix, whose either intact or defective local status as well as its composition may inhibit or stimulate migratory behavior (64). A highly impressive example for the role of the extracellular matrix in cell migration has become evident in studies on the application of stem cells in brain injury (65-71). The application of neuronal or even mesenchymal stem cells, with the original intention of replacing lost neurons, led to the remarkable finding that the transferred stem cells induced a local remodeling of the extracellular matrix, which formed a so-called biobridge that directed endogenous stem cells towards the site of injury. With regard to the numerous studies on the protective and healing effects of melatonin in various forms of brain injury (8, 72-76), it would be of considerable interest to study whether melatonin also participates in biobridge formation.

5. ROLE OF THE MELATONIN – SIRTUIN CONNECTION

The actions of melatonin are mostly explained by canonical signal transduction pathways transmitted via the MT₁ and MT₂ receptors. Even the proximate pathways prove to be complex in their details, as they contain several alternatives and are additionally modulated by receptor-binding proteins (75). Meanwhile, melatonin has been shown to influence the expression and activities of several other regulation factors, properties that expand melatonin's spectrum of effects (40, 76, 77). These factors include redox regulators, pro- and anti-inflammatory cytokines, numerous microRNAs and other noncoding RNAs, and, in particular, several

sirtuins, among which SIRT1 has been most often studied (42). Thus, melatonin's modes of signaling have been substantially broadened by the additional secondary routes. With regard to SIRT1, the relationship to melatonin is important under two aspects. First, in nontumor cells, SIRT1 is typically upregulated by melatonin, but downregulated in cancer cells (42), a surprising difference, which has been explained by the dysregulation of the sirtuin in cancer that is caused by silencing of tumor-suppressing components of cellular circadian oscillators (78). Second, melatonin and SIRT1 act, in many cases, in the same way, which has become particularly evident in the control of inflammation. Moreover, melatonin's anti-inflammatory effects were shown to be suppressed, in these cases, by sirtuin inhibitors such as sirtinol or EX527 as well as by *Sirt1* siRNA, findings that lead to the conclusion that SIRT1 transmits such actions of melatonin (20, 40, 42, 79, 80).

In addition to the effects previously summarized (20, 40, 42), other recent data on actions by sirtuins have revealed various new insights. These concern several aspects that are addressed in this review, such as programming (81, 82) and proliferation (82, 83) of stem cells, in particular, MSCs, differentiation (59), wound healing and tissue repair (58, 59, 84-87), inflammation (63, 91, 93, 94), as well as keratinocyte and fibroblast migration (63, 64, 95). Although the relationship to melatonin was only occasionally addressed in these papers (89), the overlap with melatonin's known spectrum of actions is so evident that future studies should use corresponding experimental approaches to re-investigate the possible, and presumably likely, involvement of melatonin in these systems.

Notably, the relatively strong relationship between melatonin and SIRT1 should be taken as a reason for expanding such studies to other sirtuin subforms. With regard to mitochondrial effects of melatonin, the mitochondrially located SIRT3 should be of particular interest. A few respective findings had been previously discussed (42). The importance of SIRT3 in mitochondria of stem cells has been recently addressed (91) and should be worth further efforts to connect these findings with melatonin research. Another possible connection may be assumed for melatonin and SIRT6. This subform is strongly associated with chromatin and driven in the same way by the NAD⁺ cycle as known for SIRT1 (42). In functional terms, SIRT6 was recently shown to be involved in processes that are reminiscent of actions by melatonin concerning healing of nerve injury, cell migration and M2 polarization of macrophages (92).

6. CONCLUSIONS

Melatonin's remarkable potential of protecting cells against oxidative damage and various other forms of stress has been documented in many hundreds of publications. However, defense against damage is only one side of the coin, especially as protection rarely leads to a fully complete prevention of damage. Moreover, sudden events such as traumatic injuries, ischemic or hemorrhagic insults, and injuries by accidents are poorly or not prevented by preceding cautious measures. Therefore, the healing of unavoids damage including the suppression of its undesired consequences, such as long-lasting inflammation, is likewise important for recovery to health. Although melatonin has been shown to be highly valuable in terms of reduction of post-injury damage, the aspect of tissue repair has been to date of rather secondary interest. From this author's point of view, this gap needs more attention. Moreover, it may turn out to become a promising field of melatonin research, which might have the potential for covering the entire spectrum of the health-promoting role and value of melatonin. Additionally, the aspect of tissue repair includes novel strategies of treating losses of cells by stem cell transfer. Several studies have already demonstrated the usefulness of melatonin and its downstream factor, SIRT1, in such approaches (19, 58, 59, 61, 63, 72, 83, 94-101). In the

future, the concept of biobridges (65-70) would merit additional attention in investigations on brain tissue repair.

The main aspects of this short review on preconditions of tissue repair should be regarded as interconnected phenomena, although the respective processes largely occur in a sequential order. Therefore, activation and programming of stem cells, polarization of effector cells, and controlled migration to the site of injury should ideally be investigated from an integrating point of view. Additional aspects such as self-renewal with maintenance of stemness, regulation of inflammation and of autophagy, modification of the extracellular matrix, protection against oxidative damage, downstream signaling including noncoding RNAs, sirtuins, and exosomal transmission are intertwined with the aforementioned phenomena. Some of these details concern the classic protective role of melatonin, whereas others have been rather recently added to the melatonin field. Numerous novel findings should attract the interest of researchers working on melatonin and may promise substantial insights, especially concerning wound healing, repair of the injured brain and practicability of stem cell transplantation.

On the other hand, the remarkable spectrum of protective actions should not mislead to precocious conclusions, especially by assuming that melatonin's actions are generally identical. In particular, the differences between tumor and nontumor tissues have to remain a matter of awareness. In the tumor microenvironment, actions by effector cells such as macrophages and neutrophils can be just opposite to those found in healthy tissues. This has become particularly evident concerning cell polarization, as proinflammatory phenotypes display antitumor activities. With regard to antitumor properties of melatonin, it seems to be an urgent task to clarify whether melatonin causes, in tumors, anti-inflammatory polarization as in healthy tissue or, instead, proinflammatory polarization, which would conform to its antitumor activities. In this regard, it would be also of interest to find out whether differences between tumors may exist that relate to their sensitivity to melatonin treatment.

ACKNOWLEDGMENTS

N/A

AUTHORSHIP

Manuscript and figure have been produced by R.H.

CONFLICT OF INTEREST

The author declares no conflict of interest.

REFERENCES

1. Hardeland R (2005) Antioxidative protection by melatonin – Multiplicity of mechanisms from radical detoxification to radical avoidance. *Endocrine* **27**: 119-130.
2. Hardeland R, Cardinali DP, Srinivasan V, Spence SW, Brown GM, Pandi-Perumal SR (2011) Melatonin – A pleiotropic, orchestrating regulator molecule. *Prog. Neurobiol.* **93**: 350-384.
3. Xia Y, Zeng S, Zhao Y, Zhu C, Deng B, Zhu G, *et al.* (2019) Melatonin in macrophage biology: Current understanding and future perspectives. *J. Pineal Res.* **66**: e12547.
4. Hardeland R (2018) Melatonin and inflammation —Story of a double-edged blade. *J. Pineal Res.* **65**: e12525.

5. Carrillo-Vico A, Guerrero JM, Lardone PJ, Reiter RJ (2005) A review of the multiple actions of melatonin on the immune system. *Endocrine* **27**: 189-200.
6. Hardeland R (2013) Melatonin and the theories of aging: a critical appraisal of melatonin's role in antiaging mechanisms. *J. Pineal Res.* **55**: 325-356.
7. Hardeland R, Cardinali DP, Brown GM, Pandi-Perumal SR (2015) Melatonin and brain inflammaging. *Prog. Neurobiol.* **127-128**: 46-63.
8. Reiter RJ, Mayo JC, Tan D-X, Sainz RM, Alatorre-Jimenez M, Qin L (2016) Melatonin as an antioxidant: under promises but over delivers. *J. Pineal Res.* **61**: 253-278.
9. Gensel J, Zhang B (2015) Macrophage activation and its role in repair and pathology after spinal cord injury. *Brain Res.* **1619**: 1-11.
10. Zhang Q, Sioud M. (2023) Tumor-associated macrophage subsets: shaping polarization and targeting. *Int. J. Mol. Sci.* **24**: 7493.
11. Arora S, Dev K, Agarwal B, Das P, Syed MA (2018) Macrophages: Their role, activation and polarization in pulmonary diseases. *Immunology* **223**: 383-396.
12. Hu W, Lin J, Lian X, Yu F, Liu W, Wu Y, *et al.* (2019) M2a and M2b macrophages predominate in kidney tissues and M2 subpopulations were associated with the severity of disease of IgAN patients. *Clin. Immunol.* **205**: 8-15.
13. Pugazhenthii K, Kapoor M, Clarkson AN, Hall I, Appleton I (2008) Melatonin accelerates the process of wound repair in full-thickness incisional wounds. *J. Pineal Res.* **44**: 387-396.
14. Avila-Ponce de León U, Vázquez-Jiménez A, Matadamas-Guzman M, Pelayo R, Resendis-Antonio O (2021) Transcriptional and microenvironmental landscape of macrophage transition in cancer: A Boolean analysis. *Front. Immunol.* **12**: 642842
15. Lanoix D, Lacasse AA, Reiter RJ, Vaillancourt C (2012) Melatonin: the smart killer: the human trophoblast as a model. *Mol. Cell. Endocrinol.* **348**: 1-11.
16. Slominski A, Hardeland R, Zmijewski MA, Slominski SM, Reiter RJ, Paus R (2018) Melatonin: A cutaneous perspective on its production, metabolism, and functions. *J. Invest. Dermatol.* **138**: 490-499.
17. Bjørklund G, Dadar M, Aaseth J, Chirumbolo S (2020) Thymosin β 4: a multi-faceted tissue repair stimulating protein in heart injury. *Curr. Med. Chem.* **25**: 6294-6305.
18. Liu W, Yu M, Xie D, Wang L, Ye C, Zhu Q, *et al.* (2020) Melatonin-stimulated MSC-derived exosomes improve diabetic wound healing through regulating macrophage M1 and M2 polarization by targeting the PTEN/AKT pathway. *Stem Cell Res. Ther.* **11**: 259.
19. Hardeland R (2022) Melatonin and the programming of stem cells. *Int. J. Mol. Sci.* **23**: 1971.
20. Hardeland R (2021) Melatonin and microglia. *Int. J. Mol. Sci.* **22**: 8296.
21. Reiter RJ, Sharma R, Tan D-X, Chuffa LGA, da Silva DGH, Slominski AT, *et al.* (2024) Dual sources of melatonin and evidence for different primary functions. *Front. Endocrinol. (Lausanne)* **15**: 1414463.
22. Markus RP, Cecon E, Pires-Lapa MA (2013) Immune-pineal axis: nuclear factor kappaB (NF- κ B) mediates the shift in the melatonin source from pinealocytes to immune competent cells. *Int. J. Mol. Sci.* **14**: 10979-10997.
23. Markus RP, Fernandes PA, Kinker GS, da Silveira Cruz-Machado S, Marçola M (2018) Immune-pineal axis - acute inflammatory responses coordinate melatonin synthesis by pinealocytes and phagocytes. *Br. J. Pharmacol.* **175**: 3239-3250.
24. Markus RP, Sousa KS, da Silveira Cruz-Machado S, Fernandes PA, Ferreira ZS (2021) Possible role of pineal and extra-pineal melatonin in surveillance, immunity, and first-line defense. *Int. J. Mol. Sci.* **22**: 12143.
25. Radio NM, Doctor JS, Witt-Enderby PA (2006) Melatonin enhances alkaline phosphatase activity in differentiating human adult mesenchymal stem cells grown in osteogenic

- medium via MT2 melatonin receptors and the MEK/ERK1/2 signaling cascade. *J. Pineal Res.* **40**: 332-342.
26. Luchetti F, Canonico B, Bartolini D, Arcangeletti M, Ciffolilli S, Murdolo G, *et al.* (2014) Melatonin regulates mesenchymal stem cell differentiation: a review. *J. Pineal Res.* **56**: 382-397.
 27. Lai M, Jin Z, Tang Q, Lu M (2017) Sustained release of melatonin from TiO₂ nanotubes for modulating osteogenic differentiation of mesenchymal stem cells in vitro. *J. Biomater. Sci. Polym. Ed.* **28**: 1651-1664.
 28. Dong P, Gu X, Zhu G, Li M, Ma B, Zi Y (2018) Melatonin induces osteoblastic differentiation of mesenchymal stem cells and promotes fracture healing in a rat model of femoral fracture via neuropeptide Y/neuropeptide Y receptor Y1 signaling. *Pharmacology* 2018, **102**: 272-280.
 29. Maria S, Samsonraj RM, Munmun F, Glas J, Silvestros M, Kotlarczyk MP, *et al.* (2018) Biological effects of melatonin on osteoblast/osteoclast cocultures, bone, and quality of life: Implications of a role for MT2 melatonin receptors, MEK1/2, and MEK5 in melatonin-mediated osteoblastogenesis. *J. Pineal Res.* **64**: e12465.
 30. Jiang T, Xia C, Chen X, Hu Y, Wang Y, Wu J, *et al.* (2019) Melatonin promotes the BMP9-induced osteogenic differentiation of mesenchymal stem cells by activating the AMPK/beta-catenin signalling pathway. *Stem Cell Res. Ther.* **10**: 408.
 31. Xu L, Liu Y, Sun Y, Wang B, Xiong Y, Lin W, *et al.* (2017). Tissue source determines the differentiation potentials of mesenchymal stem cells: a comparative study of human mesenchymal stem cells from bone marrow and adipose tissue. *Stem Cell Res. Ther.* **8**: 275.
 32. Wang Y, Barthez M, Chen D (2023) Mitochondrial regulation in stem cells. *Trends Cell Biol.* **2023**: S0962-8924(23)00207-6.
 33. Hardeland R, Poeggeler B, Pappolla MA (2009) Mitochondrial actions of melatonin — an endeavor to identify their adaptive and cytoprotective mechanisms. *Proc. Sax. Acad. Sci., Math. Nat. Class*, **65** (3): pp. 14-31.
 34. Acuña Castroviejo D, López LC, Escames G, López A, García JA, Reiter RJ (2011) Melatonin-mitochondria interplay in health and disease. *Curr. Top. Med. Chem.* **11**: 221-240.
 35. Hardeland R (2017) Melatonin and the electron transport chain. *Cell. Mol. Life Sci.* **74**: 3883-3896.
 36. Reiter RJ, Tan D-X, Rosales-Corral S, Galano A, Zhou XJ, Xu B (2018) Mitochondria: central organelles for melatonin's antioxidant and anti-aging actions. *Molecules* **23**: 509.
 37. Prado NJ, Ferder L, Manucha W, Diez ER (2018) Anti-inflammatory effects of melatonin in obesity and hypertension. *Curr. Hypertens. Rep.* **20**: 45.
 38. Tan D-X, Hardeland R (2020) Targeting host defense system and rescuing compromised mitochondria to increase tolerance against pathogens by melatonin may impact outcome of deadly virus infection pertinent to COVID-19. *Molecules* **25**: 4410.
 39. Stacchiotti A, Favero G, Rodella LF (2020) Impact of melatonin on skeletal muscle and exercise. *Cells* **9**: 288.
 40. Hardeland R (2018) Recent findings in melatonin research and their relevance to the CNS. *Cent. Nerv. Syst. Agents Med. Chem.* **18**: 102-114.
 41. Xu S, Li L, Wu J, An S, Fang H, *et al.* (2021) Melatonin attenuates sepsis-induced small-intestine injury by upregulating SIRT3-mediated oxidative-stress inhibition, mitochondrial protection, and autophagy induction. *Front. Immunol.* **12**: 625627.
 42. Hardeland R (2021): Sirtuins, melatonin, and the relevance of circadian oscillators. In: *Sirtuin Biology in Medicine. Targeting New Avenues of Care in Development, Aging, and Disease* (Maiese K, ed.), Academic Press, London – San Diego, CA – Cambridge, MA –

- Oxford, pp. 137-151.
43. Niu B, Li B, Wu C, Wu J, Yan Y, Shang R, *et al.* (2016) Melatonin promotes goat spermatogonia stem cells (SSCs) proliferation by stimulating glial cell line-derived neurotrophic factor (GDNF) production in Sertoli cells. *Oncotarget* **7**: 77532-77542.
 44. Bai C, Gao Y, Zhang X, Yang W, Guan W (2018) Melatonin promotes self-renewal of nestin-positive pancreatic stem cells through activation of the MT2/ERK/SMAD/nestin axis. *Artif. Cells Nanomed. Biotechnol.* **46**: 62-74.
 45. Gao Y, Ma L, Bai C, Zhang X, Yang W (2019) Melatonin promotes self-renewal and nestin expression in neural stem cells from the retina. *Histol. Histopathol.* **34**: 645-654
 46. Kwon HS, Koh SH (2020) Neuroinflammation in neurodegenerative disorders: the roles of microglia and astrocytes. *Transl. Neurodegener.* **9**: 42.
 47. Liddel SA, Barres BA (2017) Reactive astrocytes: production, function, and therapeutic potential. *Immunity* **46**: 957-967.
 48. Oksanen M, Lehtonen S, Jaronen M, Goldsteins G, Hamalainen RH, Koistinaho J (2019) Astrocyte alterations in neurodegenerative pathologies and their modeling in human induced pluripotent stem cell platforms. *Cell. Mol. Life Sci.* **76**: 2739-2760.
 49. Tiwari-Woodruff S, Morales LB, Lee R, Voskuhl RR (2007) Differential neuroprotective and antiinflammatory effects of estrogen receptor (ER) alpha and ERbeta ligand treatment. *Proc. Natl. Acad. Sci. USA* **104**: 14813-14818.
 50. Metcalfe S, Anselmi N, Escobar A, Visser MB, Kay JG (2021) Innate phagocyte polarization in the oral cavity. *Front. Immunol.* **12**: 768479.
 51. Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, *et al.* (2009) Polarization of tumor-associated neutrophil phenotype by TGF- β : "N1" versus "N2" TAN. *Cancer Cell* **16**: 183-194.
 52. Yu T, Tang Q, Chen X, Fan W, Zhou Z, Huang W, *et al.* (2021) TGF- β 1 and IL-17A mediate the protumor phenotype of neutrophils to regulate the epithelial-mesenchymal transition in oral squamous cell carcinoma. *J. Oral Pathol. Med.* **50**: 383-361.
 53. Kieler M, Hofmann M, Schabbauer G (2021) More than just protein building blocks: how amino acids and related metabolic pathways fuel macrophage polarization. *FEBS J.* **288**: 3694-3714.
 54. Chen H, Yang W, Xue X, Li Y, Jin Z, Ji Z (2022) Integrated analysis revealed an inflammatory cancer-associated fibroblast-based subtypes with promising implications in predicting the prognosis and immunotherapeutic response of bladder cancer patients. *Int. J. Mol. Sci.* **23**: 15970.
 55. Ramakrishnan S (2022) HIF-2 in cancer-associated fibroblasts polarizes macrophages and creates an immunosuppressive tumor microenvironment in pancreatic cancer. *Gastroenterology* **162**: 1835-1837.
 56. Ledoult E, Jendoubi M, Collet A, Guerrier T, Largy A, Speca S, *et al.* (2022) Simple gene signature to assess murine fibroblast polarization. *Sci. Rep.* **12**: 11748.
 57. Schneider MR, Schmidt-Ullrich R, Paus R (2009) The hair follicle as a dynamic miniorgan. *Curr. Biol.* **19**: R132-R142.
 58. Qiang L, Sample A, Liu H, Wu X, He Y-Y (2017) Epidermal SIRT1 regulates inflammation, cell migration, and wound healing. *Sci. Rep.* **7**: 14110.
 59. Chen X, Tong G, Fan J, Shen Y, Wang N, Gong W, *et al.* (2022) FGF21 promotes migration and differentiation of epidermal cells during wound healing via SIRT1-dependent autophagy. *Br. J. Pharmacol.* **179**: 1102-1121.
 60. Zhu R, Liu C, Gundersen GG (2018) Nuclear positioning in migrating fibroblasts. *Semin. Cell Dev. Biol.* **82**: 41-50.
 61. Patschan D, Hildebrandt A, Rinneburger J, Wessels JT, Patschan S, Becker JU, *et al.* (2012) The hormone melatonin stimulates renoprotective effects of "early outgrowth"

- endothelial progenitor cells in acute ischemic kidney injury. *Am. J. Physiol. Renal Physiol.* **302**: F1305-F1312.
62. Zhang J, Li L, Yu J, Fan Zhang F, Shi J, Li M, *et al.* (2023) Autophagy-modulated biomaterial: A robust weapon for modulating the wound environment to promote skin wound healing. *Int. J. Nanomedicine* **18**: 2567-2588.
 63. Tan SS, Zhan W, Poon CJ, Han X, Marre D, Boodhun S, *et al.* (2018) Melatonin promotes survival of nonvascularized fat grafts and enhances the viability and migration of human adipose-derived stem cells via down-regulation of acute inflammatory cytokines. *J. Tissue Eng. Regen. Med.* **12**: 382-392.
 64. Woodley JP, Lambert DW, Ortega Asencio I (2022) Understanding fibroblast behavior in 3D biomaterials. *Tissue Eng. Part B Rev.* **28**: 569-578.
 65. Tajiri N, Kaneko Y, Shinozuka K, Ishikawa H, Yankee E, McGrogan M, *et al.* (2013) Stem cell recruitment of newly formed host cells via a successful seduction? Filling the gap between neurogenic niche and injured brain site. *PLoS One* 2013, **8**: e74857.
 66. Tajiri N, Duncan K, Antoine A, Pabon M, Acosta SA, de la Pena I, *et al.* (2014) Stem cell-paved biobridge facilitates neural repair in traumatic brain injury. *Front. Syst. Neurosci.* **8**: 116.
 67. Sullivan R, Duncan K, Dailey T, Kaneko Y, Tajiri N, Borlongan CV (2015) A possible new focus for stroke treatment - migrating stem cells. *Expert Opin. Biol. Ther.* **15**: 949-958.
 68. Duncan K, Gonzales-Portillo GS, Acosta SA, Kaneko Y, Borlongan CV, Tajiri N (2015) Stem cell-paved biobridges facilitate stem transplant and host brain cell interactions for stroke therapy. *Brain Res.* **1623**: 160-165.
 69. Crowley MG, Tajiri N (2017) Exogenous stem cells pioneer a biobridge to the advantage of host brain cells following stroke: New insights for clinical applications. *Brain Circ.* **3**: 130-134.
 70. Lee JY, Xu K, Nguyen H, Guedes VA, Borlongan CV, Acosta SA (2017) Stem cell-induced biobridges as possible tools to aid neuroreconstruction after CNS injury. *Front. Cell Dev. Biol.* **5**: 51.
 71. Liska MG, Crowley MG, Nguyen H, Borlongan CV (2017) Biobridge concept in stem cell therapy for ischemic stroke. *J. Neurosurg. Sci.* **61**: 173-179.
 72. Osier N, McGreevy E, Pham L, Puccio A, Ren D, Conley YP, *et al.* (2018) Melatonin as a therapy for traumatic brain injury: A review of published evidence. *Int. J. Mol. Sci.* **19**: 1539.
 73. Blum B, Kaushal S, Khan S, Kim JH, Alvarez Villalba CL (2021) Melatonin in Traumatic Brain Injury and Cognition. *Cureus* **13**: e17776.
 74. Li D, He T, Zhang Y, Liu J, Zhao H, Wang D, *et al.* (2023) Melatonin regulates microglial polarization and protects against ischemic stroke-induced brain injury in mice. *Exp. Neurol.* **367**: 114464.
 75. Hardeland R (2009) Melatonin: Signaling mechanisms of a pleiotropic agent. *BioFactors* **35**: 183-192.
 76. Hardeland R (2017) The expanding functions of melatonin and their consequences to signaling. In: *Mini-Reviews in Recent Melatonin Research* (Hardeland R, ed.), Cuvillier, Göttingen, pp. 26-42.
 77. Hardeland R (2018) Extended signaling by melatonin. *Cell Cell. Life Sci. J.* **3**: 000123.
 78. Hardeland R (2014) Melatonin, noncoding RNAs, messenger RNA stability and epigenetics — evidence, hints, gaps and perspectives. *Int. J. Mol. Sci.* **15**: 18221-18252.
 79. Zhao L, An R, Yang Y, Yang X, Liu H, Yue L, *et al.* (2015) Melatonin alleviates brain injury in mice subjected to cecal ligation and puncture via attenuating inflammation, apoptosis, and oxidative stress: the role of SIRT1 signaling. *J. Pineal Res.* **59**: 230-239.

80. Carloni S, Favrais G, Saliba E, Albertini MC, Chalon S, Longini M, *et al.* (2016) Melatonin modulates neonatal brain inflammation through endoplasmic reticulum stress, autophagy, and miR-34a/silent information regulator 1 pathway. *J. Pineal Res.* **61**: 370-380.
81. Shakibaei M, Shayan P, Busch F, Aldinger C, Buhrmann C, Lueders C, *et al.* (2012) Resveratrol mediated modulation of sirt-1/Runx2 promotes osteogenic differentiation of mesenchymal stem cells: potential role of Runx2 deacetylation. *PLoS ONE* **7**: e35712.
82. Denu RA, Hematti P (2016) Effects of oxidative stress on mesenchymal stem cell biology. *Oxid. Med. Cell. Longev.* **2016**: 2989076.
83. Yuan H-F, Zhai C, Yan X-L, Zhao DD, Wang J-X, Zeng Q, *et al.* (2012) SIRT1 is required for long-term growth of human mesenchymal stem cells. *J. Mol. Med.* **90**: 389-400.
84. Shi R, Jin Y, Hu W, Lian W, Cao C, Han S, *et al.* (2020) Exosomes derived from mmu_circ_0000250-modified adipose-derived mesenchymal stem cells promote wound healing in diabetic mice by inducing miR-128-3p/SIRT1-mediated autophagy. *Am. J. Physiol. Cell Physiol.* **318**: C848-C856.
85. Christovam AC, Theodoro V, Mendonça FAS, Esquisatto MAM, Dos Santos GMT, do Amaral MEC. (2019) Activators of SIRT1 in wound repair: an animal model study. *Arch. Dermatol. Res.* **311**: 193-201.
86. Zou J, Duan Y, Wang Y, Liu A, Chen Y, Guo D, *et al.* (2022) Phellopterin cream exerts an anti-inflammatory effect that facilitates diabetes-associated cutaneous wound healing via SIRT1. *Phytomedicine* **107**: 154447.
87. Beegum F, Anuranjana PV, George KT, Divya KP, Begum F, Krishnadas N, *et al.* (2022) Sirtuins as therapeutic targets for improving delayed wound healing in diabetes. *J. Drug Target* **30**: 911-926.
88. Fu Y, Wang Y, Liu Y, Tang C, Cai J, Chen G, *et al.* (2022) p53/sirtuin 1/NF-κB signaling axis in chronic inflammation and maladaptive kidney repair after cisplatin nephrotoxicity. *Front. Immunol.* **13**: 925738.
89. Perrone S, Carloni S, Dell'Orto VG, Filonzi L, Beretta V, Petrolini C, *et al.* (2023). Hypoxic ischemic brain injury: animal models reveal new mechanisms of melatonin-mediated neuroprotection. *Rev. Neurosci.* **35** (3): 331-339.
90. Zhao W, Zhang R, Zang C, Zhang L, Zhao R, Li Q, *et al.* (2022) Exosome derived from mesenchymal stem cells alleviates pathological scars by inhibiting the proliferation, migration and protein expression of fibroblasts via delivering miR-138-5p to target SIRT1. *Int. J. Nanomedicine* **17**: 4023-4038.
91. Wang Y, Barthez M, Chen D (2023) Mitochondrial regulation in stem cells. *Trends Cell Biol.* [online ahead of print, Oct 31]; doi: 10.1016/j.tcb.2023.10.003.
92. Zou Y, Zhang J, Xu J, Fu L, Xu Y, Wang X, *et al.* (2021) SIRT6 inhibition delays peripheral nerve recovery by suppressing migration, phagocytosis and M2-polarization of macrophages. *Cell. Biosci.* **11**: 210.
93. Mendivil-Perez M, Soto-Mercado V, Guerra-Librero A, Fernandez-Gil BI, Florido J, Shen YQ, *et al.* (2017) Melatonin enhances neural stem cell differentiation and engraftment by increasing mitochondrial function. *J. Pineal Res.* **63**: e12415.
94. Zhang S, Chen S, Li Y, Liu Y (2017) Melatonin as a promising agent of regulating stem cell biology and its application in disease therapy. *Pharmacol. Res.* **117**: 252-260.
95. Lee MS, Yin TC, Sung PH, Chiang JY, Sun CK, Yip HK (2017) Melatonin enhances survival and preserves functional integrity of stem cells: A review. *J. Pineal Res.* **62**: e12372.
96. Hu C, Li L (2019) Melatonin plays critical role in mesenchymal stem cell-based regenerative medicine in vitro and in vivo. *Stem Cell Res. Ther.* **10**: 13.
97. Zhao L, Hu C, Zhang P, Jiang H, Chen J (2020) Melatonin preconditioning is an effective

- strategy for mesenchymal stem cell-based therapy for kidney disease. *J. Cell. Mol. Med.* **24**: 25-33.
98. Abdel-Kawi SH, Hashem KS (2022) Administration of melatonin in diabetic retinopathy is effective and improves the efficacy of mesenchymal stem cell treatment. *Stem Cells Int.* **2022**: 6342594.
99. Fang XY, Zhao DW, Zhang C, Ge HF, Zhang XY, Zhao FC, *et al.* (2022) A three-dimensional matrix system containing melatonin and neural stem cells repairs damage from traumatic brain injury in rats. *Neural Regen. Res.* **17**: 2512-2517.
100. Qin W, Wang J, Hu Q, Qin R, Ma N, Zheng F, *et al.* (2023) Melatonin-pretreated human umbilical cord mesenchymal stem cells improved endometrium regeneration and fertility recovery through macrophage immunomodulation in rats with intrauterine adhesions. *Biol. Reprod.* **109**: 918-937.
101. Luchetti F, Carloni S, Nasoni MG, Reiter RJ, Balduini W (2023) Melatonin, tunneling nanotubes, mesenchymal cells, and tissue regeneration. *Neural Regen. Res.* **18**: 760-762.



This work is licensed under a [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/)

Please cite this paper as:

Hardeland, R. 2024. Cell polarization, migration and tissue repair: A promising field for future melatonin research. Melatonin Research. 7, 2 (Jul. 2024), 120-133. DOI:https://doi.org/https://doi.org/10.32794/mr112500171.