Review

Multiple strategies of melatonin protecting against cardiovascular injury related to inflammation: A comprehensive overview

Swaimanti Sarkar^a, Aindrila Chattopadhyay^b, Debasish Bandyopadhyay^{a*}

^aOxidative Stress and Free Radical Biology Laboratory, Department of Physiology, University of Calcutta, 92, APC Road, Kolkata-700009, India ^bDepartment of Physiology, Vidyasagar College, 39, Sankar Ghosh Lane, Kolkata-700006, India

*Correspondence: debasish63@gmail.com, dbphys@caluniv.ac, Tel: +91-9433072066

Running title: Melatonin in inflammatory cardiovascular injury

Received: August 16, 2020; Accepted: October 15, 2020

ABSTRACT

The onset and progression of baneful chronic diseases are often accompanied by a torrent of uncontrolled inflammatory reactions. Although inflammation is a natural response to detect, eliminate, and counterpoise the harmful insults to physiological integrity, a persistent inflammation causes tissue damage or more serious disorders, for example, the atherosclerosis and myocardial infarction. Inflammation often occurs in the cardiovascular system, but are also caused by other disorders including metabolic syndrome, autoimmune diseases, AIDS, and cancer that can affect the cardiac health. To effectively treat heart diseases a potent remedy is necessary which not only suppresses the inflammation but also prevents inflammation-associated cardiopathogenesis. The ubiquitous antioxidant molecule melatonin has both antiinflammatory and cardioprotective activities. Melatonin executes its anti-inflammatory activity by its antioxidant function or by targeting multiple intracellular signalling cascades such as modulating cytokine profile, blocking inflammasome activation and apoptosis. Lipid dysregulation and endothelial dysfunction that play a crucial role in the pathogenesis of atherosclerosis, insulin resistance, and diabetes are prevented by melatonin. Attenuation of mitochondrial and ER stress by melatonin is also pertinent to its cardioprotective action. Additionally, melatonin by its immuno-stimulatory activity can suppress inflammaging and immuno-senescence in HIV patients and thereby averts chronic inflammation-induced cardiovascular abnormality in these subjects. Modulation of cytokine profile and decrease in MMP-9 secretion by melatonin is beneficial in autoimmune conditions. In addition to its anti-tumour potency, melatonin can reduce chemotherapy-induced cardio-toxicity in cancer patients. This review, therefore, provides a concise summary of the currently available information appertaining to the roles of melatonin in mitigation of chronic inflammation and its effect on cardiovascular integrity.

Key words: Melatonin, cardiovascular diseases, inflammation, atherosclerosis, myocardial infarction, metabolic syndrome, autoimmune diseases, immunodeficiency, cancer.

1. INTRODUCTION

The internal homeostasis of organisms is frequently threatened by various toxic and pathogenic agents as well as the malfunctions of endogenous molecular processes. These noxious challenges can lead to inevitable tissue injury or even lethal outcome in organisms. Therefore, organisms, in course of evolution, have developed a wellintegrated defence machinery that aims to neutralize the injurious stimuli and thereby to restore functional harmony in the affected tissue. A good example is the inflammatory system. Inflammation is a healing process; however, its overreaction can lead to detrimental consequences. In some cases, the latter often outweighs the beneficial effect of this innate immune process (1).

Evidence derived from epidemiological studies indicates that the cardiovascular diseases often emerge ensuing a chronic inflammatory state (2). Various hallmarks of inflammation are often associated with menacing cardiovascular events including atherosclerosis (3), myocardial infarction, and cardiac arrest (4). The aetiology can be explained by the fact that cardiac tissue is enriched with mitochondria which are both a source as well as the victim of oxidative stress. The tremendous metabolic activity of the heart requires a huge oxygen supply yet the relatively low levels of antioxidant capacity make the heart vulnerable to oxidative stress and concomitant inflammatory injury (5). Many chronic disorders including diabetes, obesity, autoimmune diseases, and cancer are associated with sustained low-grade inflammation that adversely affects the cardiovascular system, leading to cardiomyopathies, that originate independent of the traditional cardiovascular risk factors (6).

Animals, especially, mammals, are well equipped to combat the nocuous effect of persistent inflammatory reactions. The cellular antioxidant repertoire represents a checkpoint in the trajectory of inflammation-induced pathologies. This evokes the demand for selecting a suitable antioxidant to target inflammation (7). One such endogenous molecule is melatonin (N-acetyl-5-methoxytryptamine), a potent antioxidant and a powerful anti-inflammatory agent. It protects the cells against uncontrolled inflammation by modulating both pro- and anti-inflammatory processes (8). Cardioprotective actions of this indoleamine are usually mediated by the receptorindependent mechanism, whereby the amphiphilic feature allows it to pass through the biological membrane to achieve on-site protection inside the cells (9). Melatonin also exerts its action on cardiovascular system by interacting with its receptors localized in the cardiac and endothelial cells (10-11). In addition, systemic inflammatory reactions caused by chronic illnesses are efficiently prevented by melatonin, thus arresting the imminent attack on the cardiovascular tissues (12). Melatonin not only inhibits the persistent migration of leukocytes (13) but also suppresses the production of reactive oxygen species (ROS) (14), prevents lipid oxidation and resultant lipotoxicity (15), inhibits inflammasome activation (16), up-regulates the antioxidant, anti-inflammatory (17), and anti-apoptotic genes while minimizing the release of pro-inflammatory cytokines and pro-apoptotic proteins (18). An in-depth description encompassing the modalities of inflammation and the target points of melatonin in various signalling events within the cardiovascular tissue will be discussed in this review.

2. INFLAMMATION AND INFLAMMATORY DISEASES

Inflammation is a pathophysiological response of the body to infection or injury in which the various components of the immune system co-ordinately activate a series of signalling processes and regulate the levels of mediator molecules in the host tissue

in an attempt to ameliorate tissue damage (1, 19). The non-immune and immune cells express surface receptors known as pattern recognition receptors (PRR) that have affinity for foreign agents as well as damaged self-substances such as mitochondrial DNA, cardiolipin and other structures released by the injured cells. PRR recognize the pathogens and damaged self-molecules via their pathogen associated molecular patterns (PAMP) and danger associated molecular patterns (DAMP), respectively (20, 21). Their interactions activate a series of signalling events known as inflammatory responses. Toll-like receptors (TLR) are among the most conserved and well explored member of the PRR family and are known to trigger intracellular pathways that activate a range of transcription factors such as the nuclear factor kappa-light-chain-enhancer of activated B cells (NFkB), mitogen-activated protein kinase (MAPK) and interferon regulatory factor 3 (IRF-3) (22-24). These transcription factors translocate into the nucleus to mediate gene expression of a set of pro-inflammatory and anti-inflammatory cytokines and chemokines. Tumour Necrosis Factor alpha (TNF-α) and Interleukin-1 beta subtype (IL-1 β) are the first pro-inflammatory molecule to be secreted which perpetuates the molecular cascade of inflammatory reactions resulting in activation of transcription factors and generation of other cytokines and proteins that control apoptosis (25). Other important pro-inflammatory markers including IL-1a, IL-6, IL-8, macrophage inflammatory protein $1-\alpha$ (MIP- 1α), interferons (IFN), colony stimulating factors and transforming growth factors (TGF) (26-27) are also recruited during inflammation. However, inflammatory mechanism is not solely characterized by the release of pro-inflammatory cytokines, rather anti-inflammatory molecules have a vital role in dampening of aggressive immune reactions. An impairment in the antiinflammatory response locally may culminate in a pernicious inflammatory state at the systemic level which may have detrimental results such as multi-organ dysfunction syndrome, septic shock, and mortality (25). In addition, inflammation associated oxidative stress can also activate NFkB, MAPK and JAK-STAT (Janus Kinase-Signal Transducer and Activator of Transcription) pathways to generate inflammatory cytokines and chemokines that make the situation worse (28). Trauma and oxidative stress increase the activity of some proinflammatory enzymes such as nitric oxide synthase, xanthine oxidase, cyclooxygenase, lipoxygenase, and NADH/NADPH oxidases (19, 29). All these events pave the way for the development of a plethora of inflammatory diseases such as autoimmune conditions, Acquired Immune Deficiency Syndrome (AIDS), metabolic syndrome and cancer which will be discussed below in detail.

2.1. Inflammatory autoimmune disorders.

Autoimmune diseases involve a broad range of inflammatory conditions that arise due to a breach in immune tolerance leading to ignition of immune response against self-molecules, a condition that is either limited to a specific organ or evolves a systemic disorder (30). Interestingly, basal level of auto-reactivity of the T and B cells to self-antigens is not considered as a pathological sign and is indispensable for the survival of mature T cells in the peripheral blood. However, the decreased threshold of the activation of lymphocytes at the genetic level evokes immune reactivity against selfmolecules (31). Additionally, external factors such as infection, certain xenobiotics and trauma may also induce or exacerbate autoimmune response (30). A current evidence has documented that a dysregulated inflammatory state may be associated with the pathogenesis of autoimmune disorders (32). The common autoimmune diseases characterized by chronic inflammation include rheumatoid arthritis, systemic lupus

erythematosus, systemic sclerosis, type-1 diabetes, psoriasis, inflammatory bowel disease and autoimmune thyroid conditions.

2.2. Human Immunodeficiency Virus (HIV) infection and AIDS.

Human Immunodeficiency Virus infection and AIDS is manifested by persistent immune activation that result in chronic inflammatory status allowing continuous viral replication, gradual cell death, loss of immune function and inflammation-associated degenerative diseases (33, 34). Although the emergence of combinational antiretroviral therapy has reduced AIDS-related mortality, yet prolonged viral suppressive treatment escalates the risk of non-AIDS-related morbidity including cardiovascular pathogenicity (35-36).

HIV antigens interact with CD4⁺ and CD8⁺ lymphocytes leading to profuse secretion of pro-inflammatory cytokines and chemotactic agents including IL-1β, IL-6, TNF-α, IFN-α, MIP-1α, chemokine ligands (CXCL-9, CXCL-10 and CCL-2), and cell adhesion molecules (CAM) such as ICAM and VCAM. This results in excessive activation of T cells, their decreased half-lives, exhaustion of T cells during viral encounter and apoptosis-mediated depletion of T cell pool (37-40). Further, with the progressive viremia, the optimally functional B cells, dendritic cells, and natural killer (NK) cells are compromised resulting in utmost immune system imbalance and premature immuno-senescence (41).

2.3. Metabolic syndrome.

The World Health Organization (WHO) has proposed that metabolic syndrome comprises of multiple clinical features with insulin resistance and/or diabetes mellitus being the hallmark disorder along with at least two of the following abnormalities which include hypertension, abnormal plasma lipid profile, abdominal obesity and increased urinary albumin excretion (42). Manifestation of insulin resistance in adipocytes abolishes the antilipolytic action of insulin, thereby increases circulating free fatty acids (FFA) which further amplifies insulin resistance in adipose tissue (43). Additionally, plasma FFA has deleterious impact on insulin homeostasis due to lipotoxicity-mediated pancreatic beta cell destruction (44).

The cardiovascular component of metabolic syndrome has its pathogenetic basis in the chronic inflammatory mechanisms that are instigated by insulin resistance (45). The adipose tissues are responsible for the up-regulation of pro-inflammatory pathways by secretion of inflammation promoting adipokines such as chemerin and leptin along with the release of other pro-inflammatory mediators including IL-8, monocyte chemotactic protein (MCP)-1, and C-reactive protein (CRP)(46-47). Besides, adipose tissue resident macrophages also release inflammatory molecules viz., TNF- α which phosphorylates and inactivates insulin receptor in both smooth muscle cells and adipocytes contributing to elevated FFA release into the circulatory pool (48). TNF- α is responsible for downregulation of adiponectin, an anti-inflammatory molecule produced by adipocytes that enhance insulin sensitivity (48, 49). Additionally, IL-6 secreted by the immune cells and adipose tissue up-regulates the production of fibrinogen and CRP in the liver (49,50). Increased circulatory fibrinogen concentration is associated with prothrombotic condition in patients with metabolic syndrome (49).

2.4. Cancer.

Chronic inflammation often associates to carcinogenesis of the inflamed tissue. The immuno-pathogenic facets of cancer progression are often observed from the biopsies that display a panoply of inflammatory cells in the tumour microenvironment (51). The mechanistic nexus between inflammation and cancer can be explained by two pathways- the intrinsic and the extrinsic pathways. Several mutational events engendering the activation of oncogenes, deactivation of tumour-suppressor genes, and initiate the intrinsic pathway leading to neoplastic growth and associated inflammatory changes, even in cases without any history of inflammatory disorders (52). On the other hand, extrinsic pathway operates when carcinogenesis is attributable to a pre-existing inflammatory condition. However, both the pathways are connected to a common inflammatory mechanism that involves activation of NFkB, JAK-STAT, and HIF-1a (Hypoxia-inducible factor 1α)-mediated cytokine and chemokine production (52). One crucial chemokine molecule secreted by the neoplastic tissue is the MCP which plays a significant role in attracting monocytes that transform into tumour associated macrophages (TAM) (53). Although TAM may exhibit mild anti-malignant effect by inducing IL-12, IL-2 expression and consequent natural killer cell activation, it also involves in tumour growth and metastasis which makes TAM a major culprit in cancer progression (54, 55). Experimentally, TAM have been demonstrated to produce several pro-angiogenic growth factors such as transforming growth factor- β (TGF- β), epidermal growth factor, platelet derived growth factor, vascular endothelial growth factors and their receptors along with the production of several extracellular proteases and pro-inflammatory cytokines including IL-1, IL-6, and TNF- α (56, 57). In addition to macrophages, other immune cells including neutrophils, eosinophils, mast cells, and T lymphocytes produce chemotactic molecules, pro-angiogenic factors, and matrix degrading proteases that augment neoplastic development (53).

3. HEART AND THE VASCULAR SYSTEM AS VICTIMS OF CHRONIC INFLAMMATION

Inflammation significantly contributes to the pathogenesis of atherosclerosis and other cardiovascular morbidity. Orchestration of a number of inflammatory signalling pathways forms a crucial link between atheroma formation and associated cardiovascular complications including myocardial infarction. The notion regarding the involvement of inflammatory mediators in atherogenesis is strengthened by the observation that LDL lowering drugs seem inefficient in fully impeding the atherosclerotic process (3). Additionally, therapeutic interventions targeting inflammation have shown outstanding improvement in the prognosis of patients suffering from arteriosclerotic heart disease (3). Atherogenic diet which causes lipotoxicity in the arterial tissue is considered to be an initial step in the development of atherogenic plaque. The lipids accumulating in the arterial wall are prone to prooxidation due to the fact that the arterial intima is shield to the direct exposure to blood antioxidants (58). These oxidized lipoproteins are trapped locally and phospholipids are responsible for NFkB-mediated transcription of VCAM-1 in the endothelial cells (58-61). Circulating monocytes attach to the VCAMs on the surface of the endothelial cells undergoing diapedeses along the gradient of chemokine MCP-1 (3, 62). Lymphocytes also penetrate in the sub-endothelial area in response to lymphocyte specific chemo-attractants [interferon- γ (IFN- γ)-inducible chemokines of the CXC family] (63). Monocytes assembling within the tunica intima constitute the tissue macrophages which then engulf the oxidized lipoprotein to form arterial foam cells. These foam cells release free radicals and cytokines that augment the proinflammatory response (3, 62). The lymphocytes interact with oxidized lipoproteins and heat shock proteins to produce cytokines that facilitate activation of other cells including foam cells (62). Consequently, matrix metalloproteinases (MMPs) are released from the foam cells. MMPs break down extracellular matrix proteins and weaken the fibrous covering around the plaque; thus, lead to vascular rupture. When the plaque splits open the tissue factor (factor-III) released by plaque resident leukocytes is exposed to blood to form thrombosis with the dying macrophages to constitute the central necrotic core of the atheroma (3, 62).

An atherosclerotic lesion formed in the coronary artery can often result in arterial occlusion and the consequent ischemic cardiac injury. Acute myocardial ischaemia, infarction and reperfusion injury involve the activation of innate and adaptive immune responses that promote oxidative stress, inflammation, apoptosis, and transient or permanent loss of cardiac function (64, 65). Recruitment of inflammatory factors is indispensable at the onset of acute cardiac infarction. These factors can aid in the removal of dead cells and cellular debris from the site of infarction (66, 67). The necrotic cardiomyocytes, fibroblasts, interstitial cells and the endothelial cells trigger innate immune response by releasing damaged DNA, proteins, and lipids. These substances are then recognized as DAMP by the TLR receptors expressed by cardiomyocytes, endothelial cells, and the immune cells (68). This culminates in the activation of NFkB signalling and upregulation of several other cytokine and chemokine molecules (69,70). The chemotactic signals contribute to the directed migration of leukocytes to the site of infarction. DAMPS can further activate the complement cascade and inflammasomes in the infarct area (70, 71). This triggers the release of pro-inflammatory cytokines including IL-1, IL-6 and IL-18 and instigates pyroptosis of cardiomyocytes (70). Further, ROS generated from dysfunctional mitochondria of the infarct zone exacerbates the inflammatory process through activation of complement pathway and secretion of chemokines and cell adhesion molecules (68).

Strong evidence has demonstrated that chronic inflammation, such as rheumatoid arthritis, is associated with carotid arteriosclerosis, and increased risk of myocardial infarction and strokes even in patients without conventional cardiovascular risk factors (72-74). Several inflammatory cytokines including IL-1 β , IL-6, IL-7, and TNF- α are hallmarks of atherosclerotic conditions to be detected in the plasma of animals with rheumatoid arthritis (75, 76). Activation of inflammatory cascades which contributes to the immuno-pathogenesis of systemic lupus erythematosus also promotes cardiovascular complications including dyslipidaemia and atherosclerosis, peripheral arterial occlusion, coronary artery disease, and stroke (77-79). Cardiac dysfunction with impaired ventricular contractility and altered diastolic function along with greater incidence of myocardial infarction have been observed in systemic sclerosis patients (80). Inflammatory bowel disease (IBD) which includes two chronic inflammatory conditions of the gut- Crohn's disease and ulcerative colitis can predispose individuals to atherosclerotic cardiovascular disease (ASCVD) (81, 82). Clinical markers of IBD including CRP, IL-1, TNF-α, anti-neutrophil cytoplasmic antibodies, IgM and IgG antibodies, and vascular endothelial growth factors attribute to leukocyte migration, ROS generation, and endothelial injury, all contribute to atherogenic cardiomyopathies (83). Cardiovascular component of metabolic syndrome is often caused by inflammation and neuro-hormonal misbalance (49). Increased serum CRP and IL-6 cytokine levels observed in metabolic syndrome is correlated with ASCVD Melatonin Research (Melatonin Res.) http://www.melatonin-research.net

(84). Besides dyslipidaemia, insulin resistance can induce hypertension by increasing serum viscosity, circulatory fibrinogen concentration and angiotensin-II production (49). Angiotensin-II has been demonstrated to augment NOX-mediated free radical generation leading to NFkB activation, platelet aggregation, endothelial dysfunction and oxidation of LDL, thereby mediating initiation and exacerbation of ASCVD (49, 85). Other chronic inflammatory diseases including cancer (86) and AIDS (87) have also been found to promote progressive atherosclerotic event and adverse cardiovascular co-morbidity.

4. THE **BUILT-IN DEFENCE** STRATEGIES AGAINST **CHRONIC INFLAMMATION**

A loss of control over progressive inflammation is potentially inimical to the systemic homeostasis. A concatenation of inflammatory events that if not resolved will gradually progress towards a chronic inflammation with irreversible organ damage (88). Fortunately, our body has developed self-limiting mechanism to curb its progress with endogenous anti-inflammatory molecules and immuno-resolvents (89, 90). From the histological perspective, an inflammation resolution phase commences from the point of extreme neutrophil invasion in the inflammatory zone and continues until all the infiltrates being eliminated from this area (90). This is achieved by the accumulated leukocytes undergoing apoptotic elimination or phagocytosis or flushing awav through systemic recirculation (90). The synthesis of pro-inflammatory molecules such as cytokines, prostaglandins, leukotrienes, CAM are down-regulated and their catabolic degradation promotes resolution process (90, 91).

Several natural molecules are actively produced in aiding termination of inflammation at the induction of resolution phase. These include the resolvins, lipoxins, protectins and maresins (92). Resolvins are endogenously synthesized lipid proresolutive mediators derived from dietary ω -3 fatty acids— eicosapentaenoic and docosahexaenoic acids (93). The pro-resolutive activity of resolvins are attributable to their ability to inhibit neutrophils, monocytes, and dendritic cell migration and downregulation of IL-1 β , TNF- α , P-selectin, and VEGF gene expression (91-94). Resolvins also enhance tissue repair and regeneration after the termination of inflammation (91). Unlike resolvins, lipoxins are synthesized from the non-dietary endogenous fatty acid— arachidonic acid (95). Lipoxins potentially inhibits neutrophil mobilization and promote recruitment of non-inflammatory macrophages that are important for phagocytosis of apoptotic neutrophil and other cellular debris (a process called efferocytosis) (96). Another vital lipid immuno-resolvent is protectin which besides promoting efferocytosis and preventing polymorphonuclear neutrophil infiltration, checks T cell recruitment and stimulates their apoptosis via TNF- α and IFN- γ signalling mechanism (91).

MELATONIN AS AN ANTI-INFLAMMATORY AGENT 5.

Melatonin is a phylogenetically old molecule, first discovered in the bovine pineal gland, chemically identified as N-acetyl-5-methoxytryptamine (97-98). From bacteria to the most advanced species, Homo sapiens, melatonin's omnipresence has intrigued the scientific world to investigate its biological functions across the species. The free radical scavenging activity of melatonin is considered to be its most ancient function that has probably made this tryptophan derivative a life sustaining molecule in most of the organisms (99). The other functions of melatonin including regulation of circadian rhythm (9, 14, 100), stimulation of antioxidant enzymes (101-102), participation in immuno-regulation (103) and maintenance of metabolic homeostasis of cells are acquired during evolution. The immuno-modulatory action of melatonin encompasses both pro-inflammatory and anti-inflammatory mechanisms (104). The pro-inflammatory property is known to play a significant role in coping with pathogenic insult, while the anti-inflammatory function is the most crucial for tissue injury prevention and recuperation.

Free radical production and inflammation are reciprocally connected process . The antioxidative and ROS scavenging property of melatonin is, therefore, critical to the anti-inflammatory activity (13, 17, 105). Melatonin directly protects the proteins, lipids, and nucleic acids from oxidative injury and also stimulates the gene expression and activities of the antioxidant enzymes to restore cell's innate ability to suppress oxidative stress (14-15, 106). The progression of inflammation is always accompanied by acute ROS production that potentiate pathogen induced tissue damage (107). For instance, in one hand, the infiltrated neutrophils trigger NADPH oxidase (NOX)-induced superoxide generation known as "respiratory burst" to kill pathogens. On the other hand, this process damages the host tissues by extensive oxidative stress (108). Melatonin can proficiently suppress NOX-mediated uncontrolled oxidative burst by the phagocytes and microglial cells; therefore, prevents an impending aggressive and chronic inflammatory state (104). ROS facilitates the intercellular communications of endothelial cells by decreasing occludin expression (109). Adhesion molecules including selectins (E- selectin and P- selectin) and cell adhesion molecules (VCAM and ICAM) expression on the vascular endothelial cells are modulated by free radicals which enhance leukocyte transmigration to the site of inflammation, a process that is attenuated by superoxide dismutase (SOD) activation (110-111), while melatonin stimulates SOD activity in tissues (112). Additionally, melatonin limits adhesion molecule expression on activated endothelial cells; thus, it prevents sustained immune cell recruitment and reduces the inflammation (13).

ROS, pathogenic substances like lipopolysaccharides (LPS), and certain inflammatory mediators activate NFkB, which in turn, stimulates the expression of genes of pro-inflammatory cytokines as well as proinflammatory enzymes, viz., cycloxygenase-2 (COX-2) and inducible nitric oxide synthase (iNOS) (113-116). Melatonin suppresses the NFkB activation; therefore, downregulates pro-inflammatory gene expression and their downstream signalling (104). For example, in LPS challenged murine cells, the NFkB-upregulated iNOS and/or COX expression was significantly attenuated by melatonin (13). In SAMP8 mice melatonin suppresses NFkB activity and reduces hepatic inflammation (117). Suppression of NFkB to inhibit inflammatory damages by melatonin in ischaemia/reperfusion injury (118-120), exercise stress (121-122), Alzheimer's disease (123), pulmonary inflammation (124), IBD (125), diabetic conditions (126), and cancer (127) has been well documented.

Melatonin reduces the pro-inflammatory cytokines, TNF- α , IFN- γ and IL-12, but increases anti-inflammatory cytokine IL-10 in LPS stimulated mice (17). Similar results have been observed in animals exposed to heat stress or ageing, where IL-10 protein expression was restored by melatonin administration (128-131). In senescent animals, melatonin minimizes expressions of pro-inflammatory cytokines- IL-1β, IL-6 and TNF- α (129-130). Melatonin also normalizes the cytokine profiles in animals subjected to strenuous exercise (121, 122) and chemical injury (132-134). It also attenuates NLRP3 inflammasome activation and apoptosis (18).

6. ROLE OF MELATONIN AS AN EFFICIENT CARDIOPROTECTIVE **MOLECULE**

Melatonin has profound beneficial effects on cardiovascular disorders. Myriad of molecular mechanisms have been proposed for its cardioprotective effect. One of them is the regulation of Sirtuin 1 (SIRT1), an NAD+ dependent class III histone deacetylase and a sensor for metabolic and inflammatory stress (135). Under oxidative stress, SIRT1 deacetylates a transcription factor called Forkhead Box-O (FOXO) and triggers the transcription of its target genes of anti-oxidant enzymes (catalase and SOD), antiapoptotic factors, and proteins that control the cell cycle (136). SIRT-1 activation is essential for cell survival and stress resistance; however, it is often down-regulated in disorders including age-related cardiac abnormalities many cardiac and ischaemia/reperfusion (I/R)-induced cardiac injuries (137). In I/R injury, melatonin consequently reduces expression and malondialdehvde upregulates SIRT1 concentration, superoxide formation and NOX, caspase-3, and BAX protein expression while increasing Bcl-2 production (135). SIRT1 expression is up-regulated by melatonin in rats treated with high fat diet and streptozotocin or subjected to I/R stress (138).

In addition to SIRT1, the Notch/Hes/PI3K/Akt signalling pathway is also involved in myocardial remodelling, regeneration and restoration. Notch-1 is critical for cellular communication and is responsible for cell proliferation, differentiation, and survival (139). Upon ligand-receptor interaction, Notch intracellular domain (NICD) is split apart from the receptor by the action of γ -secretase, and is translocated to the nucleus to induce the transcription of hairy and enhancer of split-1 (HES1) mRNA (140). Protein kinase-B (also known as Akt) is an efficient suppressor of cell death and it is activated upon docking at phosphoinositides, phosphorylated by phosphoinositide 3-kinase (PI3K). This PI3K/Akt signalling is inhibited by phosphatase and tensin homolog (PTEN), and HES1 protein is known to abolish such pathway (141). Melatonin stimulates Notch1, NICD, and HES1 production, while inhibits PTEN expression. Notch/HES1/Akt signalling up-regulates anti-apoptotic BCL-2 gene and decreases the expression of pro-apoptotic genes such as caspase-3 and BAX and this pathway is enhanced by melatonin (142). Under a stressful condition such as ischaemia, the Notch signalling serves as a cardioprotective response and melatonin enhances this signalling pathway potentiating a protective mechanism (142).

Endoplasmic reticulum (ER) stress response is implicated in functional impairment of cardiac tissue. Defective protein folding and disrupted calcium homeostasis can culminate in ER stress which is characterized by aggregation of scrambled proteins. This activates the "unfolded protein response" (UPR) with resultant protein kinase RNA-like ER kinase (PERK)-mediated phosphorylation of eukaryotic initiation factor- 2α (eIF2 α). eIF2 α , then, facilitates translocation of active transcription factor-4 (ATF4) into the nucleus to trigger transcription of mRNAs associated with autophagy and apoptosis (143). Melatonin attenuates PERK/eIF2a/ATF4 signalling-stimulated myocardial ER stress in I/R injury, possibly by the activation of pro-survival mechanisms-reperfusion injury salvage kinase (RISK) pathway and survivor activating factor enhancement (SAFE) pathway (144). Hypoxia-induced I/R damage can result in disruption of calcium homeostasis in cardiomyocyte sarcoplasmic reticulum. Melatonin efficiently extenuates I/R-induced calcium imbalance by regulating the calcium-handling proteins- sarco/endoplasmic reticulum calcium ATPase (SERCA) and sodium calcium exchanger (NCX), and enzymes such as endothelial nitric oxide synthase and calcium/calmodulin-dependent protein kinase II

(CaMKII) (145). The cardioprotective effects of melatonin is also confirmed by another way, i.e., melatonin deficiency. Ganglionectomy which causes melatonin deficiency and diminishes the expressions of melatonin receptors and SERCA pump in cardiomyocytes, has been found to augment ventricular tachycardia in rat heart subjected to I/R injury (146).

Melatonin exerts its anti-hypertensive effect by receptor-mediated activation of the anterior hypothalamic area, vascular smooth muscle relaxation, antioxidant action, and lowering the blood catecholamine (147). Melatonin regulates vascular integrity by restoring a normal mitochondrial function, modulating mitochondrial dynamics through mitofusin-2 and inhibition of mitochondrial permeability transition pore (mPTP) opening (148).

Obesity and insulin resistance are common risk factors for cardiovascular conditions. A negative correlation has been observed between serum melatonin and obesity (149). In fact, melatonin can mimic the actions of insulin and leptin in regulation of energy homeostasis via a common signalling mechanism involving PI3K and STAT-3 (149). In diabetic rats, melatonin supplementation dose-dependently raised circulatory adiponectin levels, decreased glucose intolerance and enhanced insulin sensitivity (150). Besides, melatonin can prevent platelet aggregation, curbs plasma level of cholesterol, decreases endothelial permeability and attenuates inflammatory reactions, thereby impeding atherosclerotic cardiovascular disorder (151).

7. MELATONIN SHIELDS THE CARDIOVASCULAR SYSTEM AGAINST INFLAMMATORY DISEASE-MEDIATED ADVERSE REACTIONS

The potent anti-inflammatory and cardioprotective properties of melatonin contribute to its efficiency in alleviating cardiovascular disease associated with inflammation.

7.1. Melatonin in atherosclerotic cardiovascular disease.

Aggregation of oxidized phospholipids and low-density lipoproteins in the arterial intima instigates NFkB-induced expression of CAMs in endothelial cells. This is a primary step in atherogenesis. This process promotes leukocyte recruitment and their trans-endothelial migration as mentioned previously. All these events can be minimized by melatonin. In addition, melatonin down-regulates the TLR4/myeloid differentiation primary response protein (MyD88)/NFkB signalling event, further strengthening its anti-inflammatory function (152). The oxidized LDL acts as a ligand for endothelial TLR4, melatonin reduces this ligand of TLR4. The atherosclerosis related serum high density lipoprotein, triglycerides, TNF-a, IL-6, high-sensitivity Creactive protein were all reduced by melatonin along with the reduction in foam cell count (152). Melatonin stimulates SIRT3/FOXO3/Parkin system-induced suppression of NLRP3 inflammasome, thus minimizing atherosclerotic progression (153). MMP is an important factor that is responsible for the rupture of atheromatous plaque and exacerbation of atherosclerotic condition. The MMP9 activity is inhibited by melatonin that docks at the active site of the enzyme (154).

7.2. Melatonin and myocardial infarction.

Ischaemia/reperfusion injury and myocardial infarction are manifested by aggressive inflammatory reactions that can cause severe consequences, even mortality (155). In a murine model, administration of melatonin prior to ischaemic insult attenuates TLR4 pathways by the activation of SAFE mechanism (156). Besides, melatonin-mediated regulation of several intracellular signalling cascades such as JAK-STAT MAPK and pathways, well as SIRT1/FOXO1, as Notch/Hes/PI3K/Akt/SIRT3, AMP-dependent protein kinase (AMPK)/peroxisome proliferator-activated receptor γ co-activator 1 α (PGC1 α)/SIRT3 and AMPK/Protein kinase G-1a (PKG1a)/NF-E2-related factor2 (Nrf2) axes have been implicated in defending against inflammatory responses during myocardial infarction (155, 157). Due to the lipophilic property, melatonin easily reaches cytosol and activates SIRT1, SIRT3 and Nrf2 to exert its anti-inflammatory activity (155). When melatonin is used as adjunctive therapy with primary percutaneous coronary intervention (pPCI) in acute myocardial infarction patients with ST-segment elevation, it significantly improves the efficacy of pPCI and leads to the reduction of infarct size (158,159). Although both melatonin receptor 1 (MT1) and melatonin receptor 2 (MT2) are found in the mammalian heart, cardioprotective action of melatonin in I/R injury is primarily mediated by the MT2 (160). Further, melatonin administration into hypothalamic paraventricular nucleus (PVN) has resulted in reduced level of free radicals, improvement of antioxidant activity, increase in IL-10 and decrease in NF-KB and IL-1βlevels in PVN, all these are beneficial in ameliorating inflammatory cardiac damage induced by myocardial I/R injury (161).

7.3. Melatonin in diabetic cardiomyopathy.

Type-II diabetes *mellitus* is considered to be an independent hazard to coronary heart disease and myocardial infarction (162). c-Jun NH2-terminal kinase (JNK) is one of the components of MAPK pathway to involve in post-ischaemic injury in diabetic mice model. Melatonin abrogates JNK/p53 signal-induced cardiac fibrosis and apoptosis of cardiomyocytes caused by high lipid/high glucose and hypoxic assaults (163). Melatonin was able to inhibit mitochondrial and ER stress-induced cardiac cell death in diabetes by preventing the activation of tyrosine-protein kinase or Syk, thereby, improving mitochondrial complex I activity, and repressing ROS generation, proinflammatory cytokine (TNF-a, TGF-B, and IL-6) release, SERCA peroxidation, and release of pro-apoptotic caspase-9 and caspase-12 (164). Melatonin treatment has further shown an enhancement of SOD, glutathione peroxidase, catalase activities and decrease in expression of mammalian target of rapamycin (mTOR) protein in diabetic heart. mTOR is known to be involved in the pathogenesis of type-II diabetes-induced cardiac disorders (165). Additionally, in hyperglycaemic conditions, melatonin attenuates cardiac NLRP3 inflammasome activation and the concomitant rise in inflammatory cytokines— IL-18 and IL-1 β driven by procaspase-1 cleavage, thus preventing inflammation-mediated diabetic cardiomyopathy (166). Activated NLRP3 participates in TGF-B/Smad signalling pathway in cardiac fibroblasts and consequently increases the synthesis of extracellular matrix proteins that paves the way for cardiac fibrosis. Melatonin effectively prevents such fibrotic changes in cardiac tissue by inhibiting the TGF- β /Smad pathway activation (166).

7.4. Melatonin in chronic autoimmune disease-induced cardiovascular diseases.

Several autoimmune diseases entail common inflammatory mechanisms which often cause cardiovascular disorders. Chronic inflammatory autoimmune conditions can predispose patients towards lethal cardio-pathogenic alterations even in absence of

conventional cardiovascular risk factors (74, 78, 80, 82). Although the reports regarding effects of melatonin on autoimmune inflammation are not consistent, its potential beneficial effects in this condition have drawn a great attention recently. For example, recent evidence has demonstrated that melatonin down-regulates MMP9 activity, as well as IL-1 β and TNF- α expressions in the synovial fibroblasts of patients with rheumatoid arthritis (RA) (167). Melatonin also stimulates microRNA (miR-3150a-3p) to produce anti-apoptotic action in RA patients (167,168). Both systemic lupus erythematosus (SLE) and systemic sclerosis (SS) kindle pro-inflammatory responses leading to atherogenesis and myocardial infarction (79,80), which can be targeted by melatonin (152,153, 155). The potency of melatonin in the regulation of lipid homeostasis and suppression of inflammatory pathways allows it to be a suitable candidate adjuvant to conventional therapies for cardiovascular co-morbidities in patients with systemic autoimmune conditions. Mechanistically, melatonin can switch T-helper 1(TH1) cell subset towards T-helper 2(TH2) and minimizes pro-inflammatory cytokine release, thereby ameliorating the severity of autoimmune diabetes (170). The anti-inflammatory, antioxidant, and anti-apoptotic effects of melatonin are also observed in patients with inflammatory bowel disease (IBD). Rise in circulatory TNF- α and CRP levels in IBD promotes atherogenesis (83). Melatonin significantly reduces TNF-α production in animal model of colitis and reinstate the CRP levels to its physiological range in patients with IBD (171,172), further suggesting melatonin's beneficial role in ulcerative colitis, thus, melatonin has the potential to retard IBDassociated cardiomyopathy.

7.5. Effects of melatonin on immunodeficiency-mediated impaired cardiac homeostasis.

Many studies have demonstrated an association between chronic HIV infection and thickening of the carotid artery wall (173), myocardial inflammation (174), and coronary atherosclerosis (87). The morbidity and mortality related to cardiovascular diseases are higher in HIV infected patients compared to the controls. The patients are often manifested by sustained T-cell activation, persistently escalated circulatory cytokines, gradual deterioration in immune system function, chronic inflammation, viral co-infection secondary to HIV, and adverse effects of long-term combinational anti-retroviral therapy (41). Incidence of acute myocardial infarction and stroke in patients suffering from AIDS have raised serious concern. In this regard, the antiinflammatory and immuno-stimulatory functions of melatonin could be beneficial. Further, Highly Active Antiretroviral Therapy (HAART) used for HIV treatment has been reported to induce metabolic syndrome, which is a potent risk factor for cardiovascular diseases (175). In a recent study, it has been reported that melatonin administration for a month, in HIV patients receiving HAART, reduced blood glucose level by 23%. Additionally, hypercholesterolaemia and high plasma triglycerides in patients subjected to HAART were efficiently alleviated by melatonin treatment (176).

7.6. Melatonin protects the cardiac function in cancer patients.

Several in vivo and in vitro investigations have revealed the potent oncostatic effect of melatonin (177). The fundamental mechanisms governing such anti-cancer activities of melatonin involve its antioxidant and anti-inflammatory properties. In addition, the regulation of genomic instability, modulation of tumour metabolism, induction of cancer cell apoptosis, inhibition of angiogenesis as well as epithelial-tomesenchymal transition further strengthen the anticancer effects of melatonin (178). Melatonin efficiently shields the DNA against damage and mutagenesis by stimulating the activities of cellular antioxidant molecules, inhibiting the pro-oxidant enzymes and maintaining proper functioning of mitochondrial electron transport chain (177). Melatonin improves rectification of faulty DNA replication in colon cancer (HCT-15) and breast cancer (MCF-7) cell lines (179). DNA distortion caused by UV (180), ionizing radiation (181), nucleotoxic agents such as hydrogen peroxide (182), formaldehyde (183), bisphenol A (184) and phenytoin sodium (185) has also been prevented by melatonin treatment. It is suggested that high nocturnal melatonin concentration is responsible for maintaining non-malignant phenotype in cancer cells by melatonin's ability to regulate pyruvate dehydrogenase complex/pyruvate dehydrogenase kinase axis (186). Further, melatonin halts cancer development and aggression by attenuating NFkB activation (187), destabilizing hypoxia inducible factors (188), repressing cyclins and cyclin dependent kinases (189), stimulating the expressions of tumour suppressor genes (viz., BRCA and p53) (190), promoting apoptosis (191) and impeding PI3K/Akt/mTOR cascade (192).

Besides the direct damaging consequences of the disease itself, the side-effects caused by chemotherapy, which is by far the mainstream treatment for cancer is a big concern. Many of these conventional chemotherapeutic agents evoke toxic reactions that actuate a secondary malignancy as well as adversely affect the vital organs and systems including the cardiovascular system (5). One such popular anticancer drug is doxorubicin, which has been reported to impart potent cardiotoxic effects by increasing the oxidative burden in heart tissue (193). Doxorubicin-induced cardiomyocyte injury, marked by altered electrophysiological property and increased circulating cardiac damage markers, is efficiently prevented by melatonin (194). The defensive mechanism of melatonin against doxorubicin-mediated myocardial damage includes a reduction in lipid oxidation, enhancement of antioxidant activities, preservation of mitochondrial integrity, prevention of DNA fragmentation and apoptosis in cardiomyocytes, modulation of serum lipid profiles and increase in cardioprotective zinc levels in plasma (5). Similar results have been obtained in cardiotoxicity caused by epirubicin, where melatonin co-administration was found to mitigate epirubicin-induced nitrosative stress in the heart (195). Trastuzumab, used as a part of adjuvant therapy in various neoplastic conditions, has noxious impact on the cardiac tissue health (196). Melatonin administration in rats significantly lowers trastuzumab-mediated oxidative stress and cardiac injury biomarkers to their basal levels (197). Taking these together, it can be concluded that melatonin plays a vital role in restoring cardiac homeostasis by ameliorating oxidative stress and inflammatory damage that occur as a ramification of cancer pathogenesis and the toxicity caused by various radio- and chemotherapeutic interventions.

8. SUMMARY AND CONCLUSION

Inflammation is considered to be both a saviour and a noxious process depending on the severity and perpetuity of the inflammatory reactions. Melatonin, a regulator of inflammatory reaction, acts via multiple signalling mechanisms to countervail this double-edged sword of inflammation. Since oxidative stress is an integral part of inflammatory process, the antioxidant function of melatonin plays a key role in impeding the overaction of the inflammatory response in many conditions. Melatonin suppresses the excessive production of chemokines and pro-inflammatory cytokines including TNF- α , IFN- γ , IL-1 β , IL-6, IL-12 and IL-18 and stimulates the production of

anti-inflammatory IL-10. In addition, melatonin decreases the expression and activities of the proinflammatory enzymes- iNOS and COX-2 and inhibits the excessive migration of immune cells to the inflammatory site by suppressing the expression of selectins and CAM in vascular endothelial cells. Specific to the myocardial tissue, melatonin modulates various molecular pathways, including the Sirtuin and Notchmediated signalling, thus, attenuating the adverse inflammatory reaction triggered by ischemic episode. Modulation of inflammasome activation, AMPK/PGC-1a or AMPK/PKG-1a, MAPK, JAK-STAT, SAFE and RISK pathways, maintenance of calcium balance, and restoration of a healthy lipid profile confers the protective effects against atherosclerosis, myocardial infarction of melatonin and diabetic cardiomyopathy. Melatonin protects the cardiac tissue during chronic systemic and organ-specific autoimmune conditions like systemic lupus erythematosus, systemic sclerosis, rheumatoid arthritis, type-1 diabetes and inflammatory bowel disease. These mechanisms are illustrated in Figure 1.

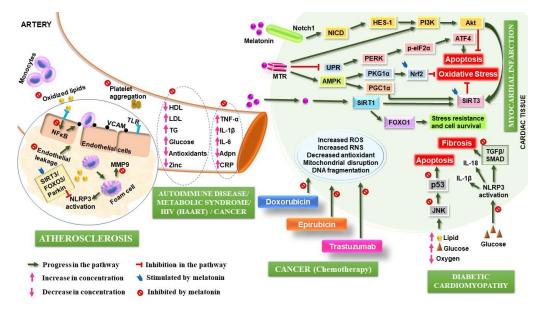


Fig. 1. Protective mechanisms of melatonin against inflammatory diseasemediated cardiovascular complications.

During atherosclerosis, oxididized LDL acts as a ligand for endothelial TLRs to activate NFk that stimulates VCAM expression in endothelial cells. Circulating monocytes adheres to the endothelial cells and increased endothelial leakage (permeability) allows their diapedesis. Within the arterial intima, monocytes are converted into tissue macrophages and by accumulating fat particles they form foam cells. Inflammatory reactions activated by foam cells and the mechanism by which melatonin prevents such responses are described in the figure. Amelioration of altered plasma composition during inflammatory conditions such as autoimmune disease, metabolic syndrome, HIV infection treated with HAART and cancer with chemotherapeutic intervention have been demonstrated. Myocardial infarction: Melatonin modulates the signaling pathways in myocardial infarction- prevents oxidative stress and apoptosis; promotes stress resistance and cell survival. Cancer (Chemotherapy): Prevention of cardiotoxic effects of doxorubicin, epirubicin and trastuzumab by melatonin has been shown. Diabetic cardiomyopathy: Attenuation of high fat, high glucose and hypoxia-induced proinflammatory pathways by melatonin in diabetic heart tissue has been demonstrated in the figure. Adpn-Adiponectin; AMPK-AMP-dependent protein kinase; ATF4- Activating transcription factor-4; CRP- C-

reactive protein; FOXO1- Forkhead Box O-1; HDL- High density lipoprotein; HES1-Hairy and enhancer of split-1; IL- Interleukin; JNK- c-Jun NH2-terminal kinase; LDL-Low density lipoprotein; MMP9- Matrix metalloproteinase-9; MTR- Melatonin receptor; NF_KB- Nuclear factor kappa-light-chain-enhancer of activated B cells; NICD1- Notch intracellular domain-1; NLRP3- NLR family, pyrin domain containing-3; Nrf2- NF-E2-related factor2; p-eIF2a- eukaryotic initiation factor-2a (phosphorylated); PERK- Protein kinase RNA-like endoplasmic reticulum kinase; PGC1a- Peroxisome proliferator-activated receptor y co-activator 1a; PI3K-Phosphoinositide 3-kinase; PKG1a- Protein kinase G-1a; ROS- Reactive oxygen species; SIRT- Sirtuin; TG- Triglycerides; TGF_β- Transforming growth factor-_β; TLR-Toll-like receptor; TNF-a- Tumor necrosis factor-a; UPR- Unfolded protein response; VCAM- Vascular cell adhesion molecule.

Undoubtedly, melatonin is a strong suppressor of excessive inflammatory reaction bestowed with versatile cardioprotective benefits. Thus, melatonin can emanate as a potent therapeutic solution for inflammatory conditions with cardiovascular pathogenicity.

AUTHORSHIP

The concept of the review article was developed by Dr. DB, Dr. AC and SS. Moreover, SS contributed in drafting the manuscript, prepared the figures, and edited it. Dr. DB and Dr. AC also revised the manuscript critically and finally approved it.

ACKNOWLEDGMENTS

Swaimanti Sarkar is extremely grateful for the financial assistance that she has received as a Junior Research fellow (JRF) [709/(CSIR-UGC NET DEC. 2018] under Joint CSIR-UGC scheme, Govt. of India. Dr. Aindrila Chattopadhyay is supported by funds available to her from Department of Science and Technology, Govt. of West Bengal. Prof. Debasish Bandyopadhyay thankfully acknowledges the support he received from Departmental BI Grant and DST-PURSE Program awarded to the University of Calcutta. Prof. DB gratefully acknowledges the critical reading and thoughtful eding of the manuscript by Dr. DunXian Tan, Editor-In-Chief, Melatonin Research. His efforts have increased definitely the scientific and readership quality of the manuscript.

CONFLICT OF INTEREST

Authors declare no conflict of interest.

REFERENCES

- 1. Okin D, Medzhitov R (2012) Evolution of inflammatory diseases. Curr. Biol. 22: R733-R740. DOI: 10.1016/j.cub.2012.07.029.
- Welsh P, Grassia G, Botha S, Sattar N, Maffia P (2017) Targeting inflammation to 2. reduce cardiovascular disease risk: a realistic clinical prospect?. Br. J. Pharmacol. 174: 3898-3913. DOI: 10.1111/bph.13818.
- Geovanini GR, Libby P (2018) Atherosclerosis and inflammation: overview and 3. updates. Clin. Sci. 132: 1243-1252. DOI: 10.1042/CS20180306.

- 4. Parekh RS, Plantinga LC, Kao WL, Meoni LA, Jaar BG, Fink NE, Powe NR, Coresh J, Klag MJ (2008) The association of sudden cardiac death with inflammation and other traditional risk factors. Kidney Int. 74: 1335-1342. DOI: 10.1038/ki.2008.449.
- 5. Ma Z, Xu L, Liu D, Zhang X, Di S, Li W, Zhang J, Reiter RJ, Han J, Li X, Yan X (2020) Utilizing melatonin to alleviate side effects of chemotherapy: a potentially good partner for treating cancer with ageing. Oxid. Med. Cell Longev. 2020: 6841581. DOI: 10.1155/2020/6841581.
- 6. Mendes AF, Cruz MT, Gualillo O (2018) The Physiology of inflammation-the common pathway to disease. Front. Physiol. 9: 1741. DOI: final 10.3389/fphys.2018.01741.
- Biswas SK (2016) Does the interdependence between oxidative stress and 7. inflammation explain the antioxidant paradox? Oxid. Med. Cell Longev. 2016: 5698931. DOI: 10.1155/2016/5698931.
- 8. Radogna F, Diederich M, Ghibelli L (2010) Melatonin: a pleiotropic molecule regulating inflammation. Biochem. Pharmacol. **80**: 1844-1852. DOI: 10.1016/j.bcp.2010.07.041.
- 9. Cipolla-Neto J, Amaral FG (2018) Melatonin as a hormone: new physiological and clinical insights. Endocr. Rev. 39: 990-1028. DOI: 10.1210/er.2018-00084.
- 10. Ekmekcioglu C, Thalhammer T, Humpeler S, Mehrabi MR, Glogar HD, Hölzenbein T, Markovic O, Leibetseder VJ, Strauss-Blasche G, Marktl W (2003) The melatonin receptor subtype MT2 is present in the human cardiovascular system. J. Pineal Res. 35: 40-44. DOI: 10.1034/j.1600-079x.2003.00051.x.
- 11. Slominski RM, Reiter RJ, Schlabritz-Loutsevitch N, Ostrom RS, Slominski AT (2012) Melatonin membrane receptors in peripheral tissues: distribution and functions. Mol. Cell. Endocrinol. 351: 152-166. DOI: 10.1016/j.mce.2012.01.004.
- 12. Nabavi SM, Nabavi SF, Sureda A, Xiao J, Dehpour AR, Shirooie S, Silva AS, Baldi A, Khan H, Daglia M (2019) Anti-inflammatory effects of Melatonin: A mechanistic review. Crit. Rev. Food Sci. Nutr. 59: S4-S16. DOI: 10.1080/10408398.2018.1487927.
- 13. Mauriz JL, Collado PS, Veneroso C, Reiter RJ, González-Gallego J (2013) A review of the molecular aspects of melatonin's anti-inflammatory actions: recent insights and new perspectives. J. Pineal Res. 54: 1-4. Doi:10.1111/j.1600-079X.2012.01014.x.
- 14. Tan DX, Reiter RJ, Manchester LC, Yan MT, El-Sawi M, Sainz RM, Mayo JC, Kohen R, Allegra MC, Hardeland R (2002) Chemical and physical properties and potential mechanisms: melatonin as a broad spectrum antioxidant and free radical scavenger. Curr. Top. Med. Chem. 2: 181-197. DOI: 10.2174/1568026023394443.
- 15. Reiter RJ, Tan DX, Kim SJ, Qi W (1998) Melatonin as a pharmacological agent against oxidative damage to lipids and DNA. Proc. West Pharmacol. Soc. 41: 229-236.
- 16. Favero G, Franceschetti L, Bonomini F, Rodella LF, Rezzani R (2017) Melatonin as an anti-inflammatory agent modulating inflammasome activation. Int. J. Endocrinol. 2017: 1835195. DOI: 10.1155/2017/1835195.
- 17. Reiter RJ, Calvo JR, Karbownik M, Qi W, Tan DX (2000) Melatonin and its relation to the immune system and inflammation. Ann. N. Y. Acad. Sci. 917: 376-386. DOI: 10.1111/j.1749-6632.2000.tb05402.x.
- 18. Carrillo-Vico A, Lardone PJ, Naji L, Fernández-Santos JM, Martín-Lacave I, Guerrero JM, Calvo JR (2005) Beneficial pleiotropic actions of melatonin in an experimental model of septic shock in mice: regulation of pro-/anti-inflammatory

cytokine network, protection against oxidative damage and anti-apoptotic effects. J. Pineal Res. 39: 400-408. DOI: 10.1111/j.1600-079X.2005.00265.x.

- 19. Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, Li Y, Wang X, Zhao L (2018) Inflammatory responses and inflammation-associated diseases in organs. Oncotarget 9: 7204–7218. DOI: 10.18632/oncotarget.23208.
- 20. Brusselle G, Bracke K (2014) Targeting immune pathways for therapy in asthma and chronic obstructive pulmonary disease. Ann. Am. Thorac. Soc. 11: S322-S328. DOI: 10.1513/AnnalsATS.201403-118AW.
- 21. Gudkov AV, Komarova EA (2016) p53 and the Carcinogenicity of chronic inflammation. Cold Spring Harb. Perspect. Med .6: a026161 DOI: 10.1101/cshperspect.a026161.
- 22. Janeway CA Jr, Medzhitov R (2002) Innate immune recognition. Annu. Rev. Immunol. 20: 197-216. DOI: 10.1146/annurev.immunol.20.083001.084359.
- 23. Yamamoto M, Takeda K (2010) Current views of toll-like receptor signaling pathways. Gastroenterol. Res. Pract. 2010: 240365. DOI: 10.1155/2010/240365.
- 24. Kawasaki T, Kawai T (2014) Toll-like receptor signaling pathways. Front. Immunol.5: 461. DOI: 10.3389/fimmu.2014.00461.
- 25. Jaffer U, Wade RG, Gourlay T (2010) Cytokines in the systemic inflammatory response syndrome: a review. HSR Proc. Intensive Care Cardiovasc. Anesth. 2: 161.
- 26. Mokart D, Capo C, Blache JL, Delpero JR, Houvenaeghel G, Martin C, Mege JL (2002) Early postoperative compensatory anti-inflammatory response syndrome is associated with septic complications after major surgical trauma in patients with cancer. Br. J. Surg. 89: 1450-1456. DOI: 10.1046/j.1365-2168.2002.02218.x.
- 27. Halter J, Steinberg J, Fink G, Lutz C, Picone A, Maybury R, Fedors N, DiRocco J, Lee HM, Nieman G (2005) Evidence of systemic cytokine release in patients undergoing cardiopulmonary bypass. J. Extra Corpor. Technol. 37: 272-277.
- 28. Reuter S, Gupta SC, Chaturvedi MM, Aggarwal BB (2010) Oxidative stress, inflammation, and cancer: how are they linked? Free Radic. Biol. Med. 49: 1603-1616. DOI: 10.1016/j.freeradbiomed.2010.09.006.
- 29. Pereira EJ, Smolko CM, Janes KA (2016) Computational models of reactive oxygen species as metabolic byproducts and signal-transduction modulators. Front. Pharmacol. 7: 457. DOI: 10.3389/fphar.2016.00457.
- 30. Smith DA, Germolec DR (1999) Introduction to immunology and autoimmunity. Environ. Health Perspect. 107: 661-665. DOI: 10.1289/ehp.99107s5661.
- 31. Yamamoto K (2004) Mechanisms of autoimmunity. Autoimmune Dis. 47: 403–406.
- 32. Rosenblum MD, Remedios KA, Abbas AK (2015) Mechanisms of human autoimmunity. J. Clin. Invest. 125: 2228-2233. DOI: 10.1172/JCI78088.
- 33. Appay V, Almeida JR, Sauce D, Autran B, Papagno L (2007) Accelerated immune senescence and HIV-1 infection. Exp. Gerontol. 42: 432-437. DOI: 10.1016/j.exger.2006.12.003.
- 34. Sereti I, Altfeld M (2016) Immune activation and HIV: an enduring relationship. Curr. Opin. HIV AIDS 11: 129-130. DOI: 10.1097/COH.00000000000244.
- 35. Hatano H (2013) Immune activation and HIV persistence: considerations for novel Curr. Opin. HIV AIDS 8: 211-216. DOI: therapeutic interventions. 10.1097/COH.0b013e32835f9788.
- 36. Miller CJ, Baker JV, Bormann AM, Erlandson KM, Hullsiek KH, Justice AC, Neuhaus J, Paredes R, Petoumenos K, Wentworth D, Winston A (2014) Adjudicated morbidity and mortality outcomes by age among individuals with HIV

infection on suppressive antiretroviral therapy. PloS One 9: e95061 DOI: 10.1371/journal.pone.0095061.

- 37. Bucy RP, Hockett RD, Derdeyn CA, Saag MS, Squires K, Sillers M, Mitsuyasu RT, Kilby JM (1999) Initial increase in blood CD4+ lymphocytes after HIV antiretroviral therapy reflects redistribution from lymphoid tissues. J. Clin. Invest. 103: 1391-1398. DOI: 10.1172/JCI5863.
- 38. Cohen Stuart JW, Hazebergh MD, Hamann D, Otto SA, Borleffs JC, Miedema F, Boucher CA, de Boer RJ (2000) The dominant source of CD4+ and CD8+ T-cell activation in HIV infection is antigenic stimulation. J. Acquir. Immune Defic. Syndr. **25**: 203-211. DOI: 10.1097/00042560-200011010-00001.
- 39. Wolf K, Tsakiris DA, Weber R, Erb P, Battegay M, Swiss HIV cohort study (2002) Antiretroviral therapy reduces markers of endothelial and coagulation activation in patients infected with human immunodeficiency virus type 1. J. Infect. Dis. 185: 456-462. DOI: 10.1086/338572.
- 40. Malherbe G, Steel HC, Cassol S, De Oliveira T, Seebregts CJ, Anderson R, Cassol E, Rossouw TM (2014) Circulating biomarkers of immune activation distinguish viral suppression from nonsuppression in HAART-treated patients with advanced HIV-1 subtype C infection. Mediators Inflamm. 2014: 198413. DOI: 10.1155/2014/198413.
- 41. Sokoya T, Steel HC, Nieuwoudt M, Rossouw TM (2017) HIV as a cause of immune activation and immunosenescence. Mediators Inflamm. 2017: 6825493. DOI: 10.1155/2017/6825493.
- 42. Alberti KG, Zimmet PZ (1998) Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus. Provisional report of a WHO consultation. Diabet. Med. 15: 539-553. DOI: https://apps.who.int/iris/handle/10665/66040.
- 43. Boden G, Shulman GI (2002) Free fatty acids in obesity and type 2 diabetes: defining their role in the development of insulin resistance and β -cell dysfunction. Eur. J. Clin. Invest. 32: 14-23.
- 44. Tooke JE, Hannemann MM (2000) Adverse endothelial function and the insulin resistance syndrome. Journal of internal medicine. 247: 425-431. DOI: 10.1046/j.1365-2362.32.s3.3.x.
- 45. Di Lorenzo C, Dell'Agli M, Colombo E, Sangiovanni E, Restani P (2013) Metabolic syndrome and inflammation: a critical review of in vitro and clinical approaches for benefit assessment of plant food supplements. Evid. Based Complement. Alternat. Med. 2013: 782461. DOI: 10.1155/2013/782461.
- 46. Barchetta I, Cimini FA, Ciccarelli G, Baroni MG, Cavallo MG (2019) Sick fat: the good and the bad of old and new circulating markers of adipose tissue inflammation. J. Endocrinol. Invest. 42: 1257-1272. DOI: 10.1007/s40618-019-01052-3.
- 47. Reddy P, Lent-Schochet D, Ramakrishnan N, McLaughlin M, Jialal I (2019) Metabolic syndrome is an inflammatory disorder: A conspiracy between adipose and phagocytes. Clin. Chim. **496**: 35-44. tissue Acta. DOI: 10.1016/j.cca.2019.06.019.
- 48. Hotamisligil GS, Murray DL, Choy LN, Spiegelman BM (1994) Tumor necrosis factor alpha inhibits signaling from the insulin receptor. Proc. Natl. Acad. Sci. 91: 4854-4858. DOI: 10.1073/pnas.91.11.4854.
- 49. Rochlani Y, Pothineni NV, Kovelamudi S, Mehta JL (2017) Metabolic syndrome: pathophysiology, management, and modulation by natural compounds. Ther. Adv. Cardiovasc. Dis. 11: 215-225. DOI: 10.1177/1753944717711379.

- 50. Fried SK, Bunkin DA, Greenberg AS (1998) Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. J. Clin. Endocrinol. Metab .83: 847-850. DOI: 10.1210/jcem.83.3.4660.
- 51. Balkwill F, Mantovani A (2001) Inflammation and cancer: back to Virchow? Lancet 357: 539-545. DOI: 10.1016/S0140-6736(00)04046-0.
- 52. Mantovani A, Allavena P, Sica A, Balkwill F (2008) Cancer-related inflammation. Nature 454: 436-444. DOI: https://doi.org/10.1038/nature07205.
- 53. Coussens LM, Werb Z (2002) Inflammation and cancer. Nature 420: 860-867. DOI: https://doi.org/10.1038/nature01322
- 54. Brigati C, Noonan DM, Albini A, Benelli R (2002) Tumors and inflammatory infiltrates: friends or foes? Clin. Exp. Metastasis 19: 247-258. DOI: 10.1023/a:1015587423262.
- 55. Tsung K, Dolan JP, Tsung YL, Norton JA (2002) Macrophages as effector cells in interleukin 12-induced T cell-dependent tumor rejection. Cancer Res. 62: 5069-5075.
- 56. Torisu H, Ono M, Kirvu H, Furue M, Ohmoto Y, Nakayama J, Nishioka Y, Sone S, Kuwano M (2000) Macrophage infiltration correlates with tumor stage and angiogenesis in human malignant melanoma: possible involvement of TNFa and 182-188. IL-1α. Int. J. Cancer **85**: DOI: 10.1002/(SICI)1097-0215(20000115)85:23.0.CO;2-M.
- 57. Schoppmann SF, Birner P, Stöckl J, Kalt R, Ullrich R, Caucig C, Kriehuber E, Nagy K, Alitalo K, Kerjaschki D (2002) Tumor-associated macrophages express lymphatic endothelial growth factors and are related to peritumoral lymphangiogenesis. Am. J. Pathol. 161: 947-956. DOI: 10.1016/S0002-9440(10)64255-1.
- 58. Collins T, Cybulsky MI (2001) NF-κB: pivotal mediator or innocent bystander in atherogenesis? J. Clin. Invest. 107: 255-264. DOI: 10.1172/JCI10373.
- 59. Khan BV, Parthasarathy SS, Alexander RW, Medford RM (1995) Modified lowdensity lipoprotein and its constituents augment cytokine-activated vascular cell adhesion molecule-1 gene expression in human vascular endothelial cells. J. Clin. Invest. 95: 1262-1270. DOI: 10.1172/JCI117776.
- 60. Palmetshofer A, Robson SC, Nehls V (1999) Lysophosphatidic acid activates nuclear factor kappa B and induces proinflammatory gene expression in endothelial cells. Thromb. Haemost. 82: 1532-1537. DOI: 10.1055/s-0037-1614867.
- 61. Dichtl W, Nilsson L, Goncalves I, Ares MP, Banfi C, Calara F, Hamsten A, Eriksson P, Nilsson J (1999) Very low-density lipoprotein activates nuclear factorendothelial kappaB in cells. Circ. Res. **84**: 1085-1094. DOI: https://doi.org/10.1161/01.RES.84.9.1085.
- 62. Libby P (2002) Inflammation in atherosclerosis. Nature 420: 868-874. DOI: https://doi.org/10.1038/nature01323.
- 63. Mach F, Sauty A, Iarossi AS, Sukhova GK, Neote K, Libby P, Luster AD (1999) Differential expression of three T lymphocyte-activating CXC chemokines by human atheroma-associated cells. J. Clin. Invest. 104: 1041-1050. DOI: 10.1172/JCI6993.
- 64. Frangogiannis NG, Smith CW, Entman ML (2002) The inflammatory response in myocardial infarction. Cardiovasc. Res. 53: 31-47. DOI: 10.1016/s0008-6363(01)00434-5.

- 65. Liu J, Wang H, Li J (2016) Inflammation and inflammatory cells in myocardial infarction and reperfusion injury: a double-edged sword. Clin. Med. Insights Cardiol. 10: 79-84. DOI: 10.4137/CMC.S33164.
- 66. Pfeffer MA, Braunwald E (1990) Ventricular remodeling after myocardial infarction. Experimental observations and clinical implications. Circulation 81: 1161-1172. DOI: 10.1161/01.cir.81.4.1161.
- 67. Opie LH, Commerford PJ, Gersh BJ, Pfeffer MA (2006) Controversies in **367**: ventricular remodelling. Lancet 356–367. DOI: 10.1016/S0140-6736(06)68074-4.
- 68. Ong SB, Hernández-Reséndiz S, Crespo-Avilan GE, Mukhametshina RT, Kwek XY, Cabrera-Fuentes HA, Hausenloy DJ (2018) Inflammation following acute myocardial infarction: multiple players, dynamic roles, and novel therapeutic opportunities. Pharmacol. Ther. 186: 73-87. DOI: 10.1016/j.pharmthera.2018.01.001.
- 69. Marchant DJ, Boyd JH, Lin DC, Granville DJ, Garmaroudi FS, McManus BM (2012) Inflammation in myocardial diseases. Circ. Res. 110: 126-144. DOI: 10.1161/CIRCRESAHA.111.243170.
- 70. van Hout GP, Arslan F, Pasterkamp G, Hoefer IE (2016) Targeting dangerassociated molecular patterns after myocardial infarction. Expert Opin. Ther. Targets 20: 223–239. DOI: 10.1517/14728222.2016.1088005.
- 71. Timmers L, Pasterkamp G, de Hoog VC, Arslan F, Appelman Y, de Kleijn DP (2012) The innate immune response in reperfused myocardium. Cardiovasc. Res. 94: 276–283. DOI: 10.1093/cvr/cvs018.
- 72. Roman MJ, Moeller E, Davis A, Paget SA, Crow MK, Lockshin MD, Sammaritano L, Devereux RB, Schwartz JE, Levine DM, Salmon JE (2006) Preclinical carotid atherosclerosis in patients with rheumatoid arthritis. Ann. Intern. Med. 144: 249-256. DOI: 10.7326/0003-4819-144-4-200602210-00006.
- 73. Gonzalez A, Maradit Kremers H, Crowson CS, Nicola PJ, Davis III JM, Therneau TM, Roger VL, Gabriel SE (2007) The widening mortality gap between rheumatoid arthritis patients and the general population. Arthritis Rheum. 56: 3583-3587. DOI: 10.1002/art.22979.
- 74. Avina-Zubieta JA, Thomas J, Sadatsafavi M, Lehman AJ, Lacaille D (2012) Risk of incident cardiovascular events in patients with rheumatoid arthritis: a metaanalysis of observational studies. Ann. Rheum. Dis. 71: 1524-1529. DOI: 10.1136/annrheumdis-2011-200726.
- 75. Devlin CM, Kuriakose G, Hirsch E, Tabas I (2002) Genetic alterations of IL-1 receptor antagonist in mice affect plasma cholesterol level and foam cell lesion size. Proc. Natl. Acad. Sci. 99: 6280-6285. DOI: 10.1073/pnas.092324399.
- 76. Ohta H, Wada H, Niwa T, Kirii H, Iwamoto N, Fujii H, Saito K, Sekikawa K, Seishima M (2005) Disruption of tumor necrosis factor- α gene diminishes the development of atherosclerosis in ApoE-deficient mice. Atherosclerosis 180: 11-17. DOI: 10.1016/j.atherosclerosis.2004.11.016.
- 77. Chuang YW, Yu MC, Lin CL, Yu TM, Shu KH, Kao CH (2015) Risk of peripheral arterial occlusive disease in patients with systemic lupus erythematosus: a nationwide population-based cohort study. Medicine 94: e2121. DOI: 10.1097/MD.00000000002121.
- 78. Holmqvist M, Simard JF, Asplund K, Arkema EV (2015) Stroke in systemic lupus erythematosus: a meta-analysis of population-based cohort studies. RMD Open 1: e000168. DOI: 10.1136/rmdopen-2015-000168.

- 79. Kay SD, Poulsen MK, Diederichsen AC, Voss A (2016) Coronary, carotid, and lower-extremity atherosclerosis and their interrelationship in Danish patients with systemic lupus erythematosus. J. Rheumatol. **43**: 315-322. DOI: 10.3899/jrheum.150488.
- 80. Meune C, Khanna D, Aboulhosn J, Avouac J, Kahan A, Furst DE, Allanore Y (2016) A right ventricular diastolic impairment is common in systemic sclerosis and is associated with other target-organ damage. Semin. Arthritis Rheum. 45: 439-445. DOI: 10.1016/j.semarthrit.2015.07.002.
- 81. Aniwan S, Pardi DS, Tremaine WJ, Loftus EV (2018) Increased risk of acute myocardial infarction and heart failure in patients with inflammatory bowel Hepatol. diseases. Clin. Gastroenterol. **16**: 1607-1615. DOI: 10.1016/j.cgh.2018.04.031.
- 82. Bigeh A, Sanchez A, Maestas C, Gulati M (2019) Inflammatory bowel disease & the risk for cardiovascular disease: does all inflammation lead to heart disease?. S1050-1738 30139-30142. Trends Cardiovasc. Med. (19): DOI: 10.1016/j.tcm.2019.10.001.
- 83. Plevy S, Silverberg MS, Lockton S, Stockfisch T, Croner L, Stachelski J, Brown M, Triggs C, Chuang E, Princen F, Singh S (2013) Combined serological, genetic, and inflammatory markers differentiate non-IBD, Crohn's disease, and ulcerative 1139–1148. colitis patients. Inflamm. Bowel Dis. **19**: DOI: 10.1097/MIB.0b013e318280b19e.
- 84. Ridker PM, Wilson PW, Grundy SM (2004) Should C-reactive protein be added to metabolic syndrome and to assessment of global cardiovascular risk?. Circulation 109: 2818–2825. DOI: 10.1161/01.CIR.0000132467.45278.59.
- 85. Mehta PK, Griendling KK (2007) Angiotensin II cell signaling: physiological and pathological effects in the cardiovascular system. Am. J. Physiol. Cell Physiol. 292: C82-C97. DOI: 10.1152/ajpcell.00287.2006.
- 86. Fitzpatrick T, Carrier M, Le Gal G (2017) Cancer, atrial fibrillation, and stroke. Thromb. Res. 155: 101-105. DOI: 10.1016/j.thromres.2017.05.006.
- 87. Lo J, Abbara S, Shturman L, Soni A, Wei J, Rocha-Filho JA, Nasir K, Grinspoon SK (2010) Increased prevalence of subclinical coronary atherosclerosis detected by coronary computed tomography angiography in HIV-infected men. AIDS 24: 243-253. DOI: 10.1097/QAD.0b013e328333ea9e.
- 88. Dinarello CA (2010) Anti-inflammatory agents: present and future. Cell 140: 935-950. DOI: 10.1016/j.cell.2010.02.043.
- 89. Henson PM (2005) Dampening inflammation. Nat. Immunol. 6: 1179-1181. DOI: 10.1038/ni1205-1179.
- 90. Serhan CN, Brain SD, Buckley CD, Gilroy DW, Haslett C, O'Neill LA, Perretti M, Rossi AG, Wallace JL (2007) Resolution of inflammation: state of the art, definitions and terms. FASEB J. 21: 325-332. DOI: 10.1096/fj.06-7227rev.
- 91. Freire MO, Van Dyke TE (2013) Natural resolution of inflammation. Periodontol. 2000.? 63: 149-164. DOI: 10.1111/prd.12034.
- 92. Bannenberg G, Serhan CN (2010) Specialized pro-resolving lipid mediators in the inflammatory response: An update. Biochim. Biophys. Acta 1801: 1260–1273. DOI: 10.1016/j.bbalip.2010.08.002.
- 93. Bannenberg GL, Chiang N, Ariel A, Arita M, Tjonahen E, Gotlinger KH, Hong S, Serhan CN (2005) Molecular circuits of resolution: formation and actions of resolvins and protectins. J. Immunol. 174: 4345-4355. DOI: 10.4049/jimmunol.174.7.4345.

- 94. Hasturk H, Kantarci A, Goguet-Surmenian E, Blackwood A, Andry C, Serhan CN, Van Dyke TE (2007) Resolvin E1 regulates inflammation at the cellular and tissue level and restores tissue homeostasis in vivo. J. Immunol. 179: 7021-7029. DOI: 10.4049/jimmunol.179.10.7021.
- 95. Samuelsson B, Dahlen SE, Lindgren JA, Rouzer CA, Serhan CN (1987) Leukotrienes and lipoxins: structures, biosynthesis, and biological effects. Science 237: 1171–1176. DOI: 10.1126/science.2820055.
- 96. Maddox JF, Colgan SP, Clish CB, Petasis NA, Fokin VV, Serhan CN (1998) Lipoxin B4 regulates human monocyte/neutrophil adherence and motility: design of stable lipoxin B4 analogs with increased biologic activity. FASEB J. 12: 487-494 DOI: 10.1096/fasebj.12.6.487.
- 97. Lerner AB, Case JD, Takahashi Y, Lee TH, Mori W (1958) Isolation of melatonin, the pineal gland factor that lightens melanocytes. J. Am. Chem. Soc .80: 2587. DOI: https://doi.org/10.1021/ja01543a060.
- 98. Lerner AB, Case JD, Heinzelmann RV (1959) Structure of melatonin. J. Am. Chem. Soc. 81: 6084-6085. DOI: https://doi.org/10.1021/ja01531a060.
- 99. Zhao D, Yu Y, Shen Y, Liu Q, Zhao Z, Sharma R, Reiter RJ (2019) Melatonin synthesis and function: evolutionary history in animals and plants. Front. Endocrinol. 10: 249. DOI: 10.3389/fendo.2019.00249.
- 100.Cardinali, D. P., Pandi-Perumal, S. R., & Niles, L. P (2008) Melatonin and its receptors: biological function in circadian sleep-wake regulation (Cambridge University Press. Cambridge), 283-314. DOI: pp 10.1017/CBO9780511541674.011.
- 101.Tan DX, Manchester LC, Hardeland R, Lopez-Burillo S, Mayo JC, Sainz RM, Reiter RJ (2003) Melatonin: a hormone, a tissue factor, an autocoid, a paracoid, and an antioxidant vitamin. J. Pineal Res. 34: 75-78. DOI: 10.1034/j.1600-079x.2003.02111.x.
- 102.Reiter RJ, Mayo JC, Tan DX, Sainz RM, Alatorre-Jimenez M, Qin L (2016) Melatonin as an antioxidant: under promises but over delivers. J. Pineal Res. 61: 253-278. DOI: 10.1111/jpi.12360.
- 103.Carrillo-Vico A, Reiter RJ, Lardone PJ, Herrera JL, Fernández-Montesinos R, Guerrero JM, Pozo D (2006) The modulatory role of melatonin on immune responsiveness. Curr. Opin. Investig. Drugs 7: 423.
- 104.Hardeland R (2018) Melatonin and inflammation—Story of a double-edged blade. J. Pineal Res. 65: e12525. DOI: 10.1111/jpi.12525.
- 105.Galano A, Tan DX, Reiter RJ (2011) Melatonin as a natural ally against oxidative stress: a physicochemical examination. J. Pineal Res. 51: 1-6.
- 106.García JJ, López-Pingarrón L, Almeida-Souza P, Tres A, Escudero P, García-Gil FA, Tan DX, Reiter RJ, Ramírez JM, Bernal-Pérez M (2014) Protective effects of melatonin in reducing oxidative stress and in preserving the fluidity of biological membranes: a review. J. Pineal Res. 56: 225-237. DOI: 10.1111/jpi.12128.
- 107.Chelombitko MA (2018) Role of Reactive Oxygen Species in Inflammation: A Minireview. Moscow Univ. Biol. Sci. Bull. **73**: 199–202. DOI: https://doi.org/10.3103/S009639251804003X.
- 108.Segal AW (2008) The function of the NADPH oxidase of phagocytes and its relationship to other NOXs in plants, invertebrates, and mammals. Int. J. Biochem. Cell 40: 604–618. DOI: 10.1016/j.biocel.2007.10.003.
- 109.Blasig IE, Bellmann C, Cording J, Del Vecchio G, Zwanziger D, Huber O, Haseloff RF (2011) Occludin protein family: Oxidative stress and reducing conditions. Antioxid. Redox. Signaling 15: 1195–1219. DOI: 10.1089/ars.2010.3542.

- 110.Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB (2008) Reactive oxygen species in inflammation and tissue injury. Antioxid. Redox. Signaling 20: 1126-1167. DOI: 10.1089/ars.2012.5149.
- 111. Laurila JP, Laatikainen LE, Castellone MD, Laukkanen MO (2009) SOD3 reduces inflammatory cell migration by regulating adhesion molecule and cytokine expression. PLoS One 4: e5786. DOI: 10.1371/journal.pone.0005786.
- 112.Liu F, Ng TB (2000) Effect of pineal indoles on activities of the antioxidant defense enzymes superoxide dismutase, catalase, and glutathione reductase, and levels of reduced and oxidized glutathione in rat tissues. Biochem. Cell Biol. 78: 447-453. DOI: 10.1139/o00-018.
- 113.Pasparakis M, Luedde T, Schmidt-Supprian M (2006) Dissection of the NF-KB signalling cascade in transgenic and knockout mice. Cell Death Differ. 13: 861-872. DOI: 10.1038/sj.cdd.4401870.
- 114.Lawrence T (2009) The nuclear factor NF-KB pathway in inflammation. Cold Spring Harb. Perspect. Biol. 1: a001651. DOI: 10.1101/cshperspect.a001651.
- 115.García-Mediavilla V, Crespo I, Collado PS, Esteller A, Sánchez-Campos S, Tuñón MJ, González-Gallego J (2007) The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. Eur. J. Pharmacol. 557: 221-229. DOI: 10.1016/j.ejphar.2006.11.014.
- 116.Hayden MS, Ghosh S (2012) NF-kB, the first quarter-century: remarkable progress and outstanding questions. Gene Dev. 26: 203-234. DOI: 10.1101/gad.183434.111.
- 117. Cuesta S, Kireev R, Forman K, García C, Escames G, Ariznavarreta C, Vara E, Tresguerres JA (2010) Melatonin improves inflammation processes in liver of senescence-accelerated prone male mice (SAMP8). Exp. Gerontol. 45: 950-956. DOI: 10.1016/j.exger.2010.08.016.
- 118.Rodríguez-Reynoso S, Leal C, Portilla E, Olivares N, Muñiz J (2001) Effect of exogenous melatonin on hepatic energetic status during ischemia/reperfusion: possible role of tumor necrosis factoralpha and nitric oxide. J. Surg. Res. 100: 141-149. DOI: 10.1006/jsre.2001.6185.
- 119.Koh PO (2008) Melatonin regulates nitric oxide synthase expression in ischemic brain injury. J. Vet. Med. Sci. 70: 747-750. DOI: 10.1292/jvms.70.747.
- 120.Li Z, Nickkholgh A, Yi X, Bruns H, Gross ML, Hoffmann K, Mohr E, Zorn M, Büchler MW, Schemmer P (2009) Melatonin protects kidney grafts from ischemia/reperfusion injury through inhibition of NF-kB and apoptosis after experimental kidney transplantation. J. Pineal Res. 46: 365-372. DOI: 10.1111/j.1600-079X.2009.00672.x.
- 121.Alonso M, Collado PS, González-Gallego J (2006) Melatonin inhibits the expression of the inducible isoform of nitric oxide synthase and nuclear factor kappa B activation in rat skeletal muscle. J. Pineal Res. 41: 8-14. DOI: 10.1111/j.1600-079X.2006.00323.x.
- 122. Veneroso C, Tuñón MJ, González-Gallego J, Collado PS (2009) Melatonin reduces cardiac inflammatory injury induced by acute exercise. J. Pineal Res. 47: 184-191. DOI: 10.1111/j.1600-079X.2009.00699.x.
- 123.Rosales-Corral SA, Acuña-Castroviejo D, Coto-Montes A, Boga JA, Manchester LC, Fuentes-Broto L, Korkmaz A, Ma S, Tan DX, Reiter RJ (2012) Alzheimer's disease: pathological mechanisms and the beneficial role of melatonin. J. Pineal Res. 52: 167–202. DOI: 10.1111/j.1600-079X.2011.00937.x.

- 124.Chen CF, Wang D, Reiter RJ, Yeh DY (2011) Oral melatonin attenuates lung inflammation and airway hyperreactivity induced by inhalation of aerosolized pancreatic fluid in rats. J. Pineal Res. 50: 46-53. DOI: 10.1111/j.1600-079X.2010.00808.x.
- 125.Cuzzocrea S, Mazzon E, Serraino I, Lepore V, Terranova ML, Ciccolo A, Caputi AP (2001) Melatonin reduces dinitrobenzene sulfonic acid-induced colitis. J. Pineal Res. 30: 1–12. DOI: 10.1034/j.1600-079x.2001.300101.x.
- 126.Negi G, Kumar A, Sharma SS (2011) Melatonin modulates neuroinflammation and oxidative stress in experimental diabetic neuropathy: effects on NF-kappaB and Nrf2 cascades. J. Pineal Res. 50: 124-131. DOI: 10.1111/j.1600-079X.2010.00821.x.
- 127. Martínez-Campa C, González A, Mediavilla MD, Alonso-González C, Alvarez-García V, Sánchez-Barceló EJ, Cos S (2009) Melatonin inhibits aromatase promoter expression by regulating cyclooxygenases expression and activity in breast cancer cells. Br. J. Cancer 101: 1613–1619. DOI: 10.1038/sj.bjc.6605336.
- 128.Kireev RA, Tresguerres AC, Garcia C, Ariznavarreta C, Vara E, Tresguerres JA (2008) Melatonin is able to prevent the liver of old castrated female rats from oxidative and pro-inflammatory damage. J. Pineal Res. 45: 394-402. DOI: 10.1111/j.1600-079X.2008.00606.x.
- 129.Forman K, Vara E, García C, Kireev R, Cuesta S, Escames G, Tresguerres JA (2011) Effect of a combined treatment with growth hormone and melatonin in the cardiological aging on male SAMP8 mice. J. Gerontol. A. Biol. Sci. Med. Sci. 66: 823-834. DOI: 10.1093/gerona/glr083.
- 130. Cuesta S, Kireev R, García C, Forman K, Escames G, Vara E, Tresguerres JA (2011) Beneficial effect of melatonin treatment on inflammation, apoptosis and oxidative stress on pancreas of a senescence accelerated mice model. Mech. Ageing Dev. 132: 573–582. DOI: 10.1016/j.mad.2011.10.005.
- 131.Lin XJ, Mei GP, Liu J, Li YL, Zuo D, Liu SJ, Zhao TB, Lin MT (2011) Therapeutic effects of melatonin on heatstroke-induced multiple organ dysfunction syndrome in rats. J. Pineal Res. 50: 436-444. DOI: 10.1111/j.1600-079X.2011.00863.x.
- 132. Tocharus J, Khonthun C, Chongthammakun S, Govitrapong P (2010) Melatonin attenuates methamphetamine-induced overexpression of pro-inflammatory cytokines in microglial cell lines. J. Pineal Res. 48: 347-352. DOI: 10.1111/j.1600-079X.2010.00761.x.
- 133.Mishra A, Paul S, Swarnakar S (2011) Downregulation of matrix metalloproteinase-9 by melatonin during prevention of alcohol-induced liver injury in mice. Biochimie 93: 854-866. DOI: 10.1016/j.biochi.2011.02.007.
- 134.Kim GD, Lee SE, Kim TH, Jin YH, Park YS, Park CS (2012) Melatonin suppresses acrolein-induced IL-8 production in human pulmonary fibroblasts. J. Pineal Res. **52**: 356–364. DOI: 10.1111/j.1600-079X.2011.00950.x.
- 135. Yu L, Sun Y, Cheng L, Jin Z, Yang Y, Zhai M, Pei H, Wang X, Zhang H, Meng Q, Zhang Y (2014) Melatonin receptor-mediated protection against myocardial ischemia/reperfusion injury: role of SIRT 1. J. Pineal Res. 57: 228-238. DOI: 10.1111/jpi.12161.
- 136.Brunet A, Sweeney LB, Sturgill JF, Chua KF, Greer PL, Lin Y, Tran H, Ross SE, Mostoslavsky R, Cohen HY, Hu LS (2004) Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. Science 303: 2011-2015. DOI: 10.1126/science.1094637.

- 137. Alcendor RR, Gao S, Zhai P, Zablocki D, Holle E, Yu X, Tian B, Wagner T, Vatner SF, Sadoshima J (2007) Sirt1 regulates aging and resistance to oxidative stress in the heart. Circ. Res. 100: 1512-1521. DOI: 10.1161/01.RES.0000267723.65696.4a.
- 138. Yu L, Liang H, Dong X, Zhao G, Jin Z, Zhai M, Yang Y, Chen W, Liu J, Yi W, Yang J (2015) Reduced silent information regulator 1 signaling exacerbates myocardial ischemia-reperfusion injury in type 2 diabetic rats and the protective effect of melatonin. J. Pineal Res. 59: 376-390. DOI: 10.1111/jpi.12269.
- 139. Yu L, Li F, Zhao G, Yang Y, Jin Z, Zhai M, Yu W, Zhao L, Chen W, Duan W, Yu S (2015) Protective effect of berberine against myocardial ischemia reperfusion injury: role of Notch1/Hes1-PTEN/Akt signaling. Apoptosis 20: 796-810. DOI: 10.1007/s10495-015-1122-4.
- 140.Miele L (2006) Notch signaling. Clin. Cancer Res. 12: 1074-1079. DOI: 10.1158/1078-0432.ccr-05-2570.
- 141.Salmena L, Carracedo A, Pandolfi PP (2008) Tenets of PTEN tumor suppression. Cell 133: 403-414. DOI: 10.1016/j.cell.2008.04.013.
- 142. Yu L, Liang H, Lu Z, Zhao G, Zhai M, Yang Y, Yang J, Yi D, Chen W, Wang X, Duan W (2015) Membrane receptor-dependent Notch1/Hes1 activation by melatonin protects against myocardial ischemia-reperfusion injury: in vivo and in vitro studies. J. Pineal Res. 59: 420-433. DOI: 10.1111/jpi.12272.
- 143.Luo B, Lin Y, Jiang S, Huang L, Yao H, Zhuang Q, Zhao R, Liu H, He C, Lin Z (2016) Endoplasmic reticulum stress eIF2 α-ATF4 pathway-mediated cyclooxygenase-2 induction regulates cadmium-induced autophagy in kidney. Cell death Dis. 7: e2251. DOI: 10.1038/cddis.2016.78.
- 144. Yu L, Li B, Zhang M, Jin Z, Duan W, Zhao G, Yang Y, Liu Z, Chen W, Wang S, Yang J (2016) Melatonin reduces PERK-eIF2a-ATF4-mediated endoplasmic reticulum stress during myocardial ischemia-reperfusion injury: role of RISK and SAFE pathways interaction. Apoptosis 21: 809-824. DOI: 10.1007/s10495-016-1246-1.
- 145. Yeung HM, Hung MW, Lau CF, Fung ML (2015) Cardioprotective effects of melatonin against myocardial injuries induced by chronic intermittent hypoxia in rats. J. Pineal Res. 58: 12-25. DOI: 10.1111/jpi.12190.
- 146.Prado NJ, Muñoz EM, Farias Altamirano LE, Aguiar F, Ponce Zumino AZ, Sánchez FJ, Miatello RM, Pueyo E, Diez ER (2020) Reperfusion arrhythmias increase after superior cervical ganglionectomy due to conduction disorders and changes in repolarization. Int. J. Mol. Sci .21: 1804. DOI: 10.3390/ijms21051804.
- 147.Xia CM, Shao CH, Xin L, Wang YR, Ding CN, Wang J, Shen LL, Li L, Cao YX, Zhu DN (2008) Effects of melatonin on blood pressure in stress-induced hypertension in rats. Clin. Exp. Pharmacol. Physiol. 35: 1258-1264. DOI: 10.1111/j.1440-1681.2008.05000.x.
- 148.Prado NJ, Ferder L, Manucha W, Diez ER (2018) Anti-inflammatory effects of melatonin in obesity and hypertension. Curr. Hypertens. Rep. 20: 45. DOI: 10.1007/s11906-018-0842-6.
- 149.Szewczyk-Golec K, Woźniak A, Reiter RJ (2015) Inter-relationships of the chronobiotic, melatonin, with leptin and adiponectin: implications for obesity. J. Pineal Res. 59: 277-291. DOI: 10.1111/jpi.12257.
- 150.de Oliveira AC, Andreotti S, Farias Tda S, Torres-Leal FL, de Proença AR, Campaña AB, de Souza AH, Sertié RA, Carpinelli AR, Cipolla-Neto J, Lima FB (2012) Metabolic disorders and adipose tissue insulin responsiveness in neonatally STZ-induced diabetic rats are improved by longterm melatonin treatment. Endocrinology 153: 2178–2188. DOI: 10.1210/en.2011-1675.

- 151.Reiter RJ, Tan DX, Paredes SD, Fuentes-Broto L (2010) Beneficial effects of melatonin in cardiovascular disease. Ann. Med. 42: 276-285. DOI: 10.3109/07853890903485748.
- 152.Hu ZP, Fang XL, Fang N, Wang XB, Qian HY, Cao Z, Cheng Y, Wang BN, Wang Y (2013) Melatonin ameliorates vascular endothelial dysfunction, inflammation, and atherosclerosis by suppressing the TLR 4/NF-kB system in high-fat-fed rabbits. J. Pineal Res. 55: 388-398. DOI: 10.1111/jpi.12085.
- 153.Ma S, Chen J, Feng J, Zhang R, Fan M, Han D, Li X, Li C, Ren J, Wang Y, Cao F (2018) Melatonin ameliorates the progression of atherosclerosis via mitophagy activation and NLRP3 inflammasome inhibition. Oxid. Med. Cell. Longev. 2018: 9286458. DOI: 10.1155/2018/9286458.
- 154.Rudra DS, Pal U, Maiti NC, Reiter RJ, Swarnakar S (2013) Melatonin inhibits matrix metalloproteinase-9 activity by binding to its active site. J. Pineal Res .54: 398-405. DOI: 10.1111/jpi.12034.
- 155.Fu Z, Jiao Y, Wang J, Zhang Y, Shen M, Reiter RJ, Xi Q and Chen Y (2020) Cardioprotective role of melatonin in acute myocardial infarction. Front. Physiol. 11: 366. DOI: 10.3389/fphys.2020.00366.
- 156.Nduhirabandi F, Lamont K, Albertyn Z, Opie LH, Lecour S (2016) Role of tolllike receptor 4 in melatonin-induced cardioprotection. J. Pineal Res. 60: 39-47. DOI: 10.1111/jpi.12286.
- 157.Sun H, Gusdon AM, Qu S (2016) Effects of melatonin on cardiovascular diseases: progress in the past year. Curr. Opin. Lipidol. 27: 408-413. DOI: 10.1097/MOL.00000000000314.
- 158.Dominguez-Rodriguez A, Abreu-Gonzalez P, Jose M, Consuegra-Sanchez L, Piccolo R, Gonzalez-Gonzalez J, Garcia-Camarero T, del Mar Garcia-Saiz M, Aldea-Perona A, Reiter RJ, Caballero-Estevez N (2017) Usefulness of early treatment with melatonin to reduce infarct size in patients with ST-segment elevation myocardial infarction receiving percutaneous coronary intervention (from the melatonin adjunct in the acute myocardial infarction treated with angioplasty trial). Am. J. Cardiol. **120**: 522-526. DOI:10.1016/j.amjcard.2017.05.018.
- 159. Dominguez-Rodriguez A, Abreu-Gonzalez P, Chen Y (2019) Cardioprotection and effects of melatonin administration on cardiac ischemia reperfusion: Insight from studies. Melatonin 100-105. clinical Res. 2: DOI: https://doi.org/https://doi.org/10.32794/mr11250024.
- 160.Han D, Wang Y, Chen J, Zhang J, Yu P, Zhang R, Li S, Tao B, Wang Y, Qiu Y, Xu M (2019) Activation of melatonin receptor 2 but not melatonin receptor 1 mediates melatonin-conferred cardioprotection against myocardial ischemia/reperfusion injury. J. Pineal Res. 67: e12571. DOI: 10.1111/jpi.12571.
- 161. Yang JB, Kang YM, Zhang C, Yu XJ, Chen WS (2019) Infusion of melatonin into the paraventricular nucleus ameliorates myocardial ischemia-reperfusion injury by regulating oxidative stress and inflammatory cytokines. J. Cardiovasc. Pharmacol. 74: 336-347. DOI: 10.1097/FJC.000000000000011.
- 162.Zheng Y, Ley SH, Hu FB (2018) Global aetiology and epidemiology of type 2 diabetes mellitus and its complications. Nat. Rev. Endocrinol.14: 88-98. DOI: 10.1038/nrendo.2017.151
- 163.Lu L, Ma J, Sun M, Wang X, Gao E, Lu L, Ren J, Yang L, Yang J (2020) Melatonin ameliorates MI-Induced cardiac remodeling and apoptosis through a JNK/p53dependent mechanism in diabetes mellitus. Oxid. Med. Cell. Longev. 2020: 1535201. DOI: 10.1155/2020/1535201.

- 164.Zhou H, Yue Y, Wang J, Ma Q, Chen Y (2018) Melatonin therapy for diabetic cardiomyopathy: a mechanism involving Syk-mitochondrial complex I-SERCA pathway. Cell. Signal 47: 88-100. DOI: 10.1016/j.cellsig.2018.03.012.
- 165.Kandemir YB, Tosun V, Güntekin Ü (2019) Melatonin protects against streptozotocin-induced diabetic cardiomyopathy through the mammalian target of rapamycin (mTOR) signaling pathway. Adv. Clin. Exp. Med. 28: 1171-1177. DOI: 10.17219/acem/103799.
- 166.Che H, Wang Y, Li H, Li Y, Sahil A, Lv J, Liu Y, Yang Z, Dong R, Xue H, Wang L (2020) Melatonin alleviates cardiac fibrosis via inhibiting lncRNA MALAT1/miR-141-mediated NLRP3 inflammasome and TGF-\u00b31/Smads signaling in diabetic cardiomyopathy. FASEB J. 34: 5282-5298. DOI: 10.1096/fj.201902692R.
- 167.MacDonald IJ, Huang CC, Liu SC, Tang CH (2020) Reconsidering the role of melatonin in rheumatoid arthritis. Int. J. Mol. Sci. 21: 2877. DOI: 10.3390/ijms21082877.
- 168. Huang CC, Chiou CH, Liu SC, Hu SL, Su CM, Tsai CH, Tang CH (2019) Melatonin attenuates TNF- α and IL-1 β expression in synovial fibroblasts and diminishes cartilage degradation: Implications for the treatment of rheumatoid arthritis. J. Pineal Res. 66: e12560. DOI: 10.1111/jpi.12560.
- 169.Conti A, Maestroni GJ (1996) Role of the pineal gland and melatonin in the development of autoimmune diabetes in non-obese diabetic mice. J. Pineal Res. **20**: 164-172. DOI: 10.1111/j.1600-079x.1996.tb00253.x.
- 170.Lin GJ, Huang SH, Chen SJ, Wang CH, Chang DM, Sytwu HK (2013) Modulation by melatonin of the pathogenesis of inflammatory autoimmune diseases. Int. J. Mol. Sci. 14: 11742-11766. DOI: 10.3390/ijms140611742.
- 171.Mazzon E, Esposito E, Crisafulli C, Riccardi L, Muià C, Di Bella P, Meli R, Cuzzocrea S (2006) Melatonin modulates signal transduction pathways and apoptosis in experimental colitis. J. Pineal Res. 41: 363-373. DOI: 10.1111/j.1600-079X.2006.00378.x.
- 172.Chojnacki C, Wisniewska-Jarosinska M, Walecka-Kapica E, Klupinska G, Jaworek J, Chojnacki J (2011) Evaluation of melatonin effectiveness in the adjuvant treatment of ulcerative colitis. J. Physiol. Pharmacol .62: 327-334.
- 173.Hulten E, Mitchell J, Scally J, Gibbs B, Villines TC (2009) HIV positivity, protease inhibitor exposure and subclinical atherosclerosis: a systematic review and metaanalysis of observational studies. Heart **95**: 1826-1835. DOI: 10.1136/hrt.2009.177774.
- 174.Luetkens JA, Doerner J, Schwarze-Zander C, Wasmuth JC, Boesecke C, Sprinkart AM, Schmeel FC, Homsi R, Gieseke J, Schild HH, Rockstroh JK (2016) Cardiac magnetic resonance reveals signs of subclinical myocardial inflammation in asymptomatic HIV-infected patients. *Circ. Cardiovasc. Imaging.* **9**: e004091. DOI: 10.1161/CIRCIMAGING.115.004091.
- 175.Narciso P, Tozzi V, D'Offizi G, de Carli G, Orchi N, Galati V, Vincenzi L, Bellagamba R, Carvelli C, Puro V (2001) Metabolic and morphologic disorders in patients treated with hightly active antiretroviral therapy since primary HIV infection. Ann. NY Acad. Sci. 946: 214-222. DOI: 10.1111/j.1749-6632.2001.tb03914.x.
- 176.Nerone FR, Messias GF, Spack Jr M, Pupulin AR (2019) Effects of melatonin on wetabolic abnormalities in HIV patients treated with antiretroviral drugs. J. Adv. Med. Pharm. Sci. 16: 1-10. DOI: 10.9734/jamps/2019/v21i130121.

- 177. Talib WH (2018) Melatonin and cancer hallmarks. *Molecules* 23: 518. DOI: 10.3390/molecules23030518.
- 178.Li Y, Li S, Zhou Y, Meng X, Zhang JJ, Xu DP, Li HB (2017) Melatonin for the prevention and treatment of cancer. Oncotarget 8: 39896-39921. DOI: 10.18632/oncotarget.16379.
- 179.Liu R, Fu A, Hoffman AE, Zheng T, Zhu Y (2013) Melatonin enhances DNA repair capacity possibly by affecting genes involved in DNA damage responsive pathways. BMC Cell Biol. 14: 1. DOI: 10.1186/1471-2121-14-1.
- 180. Fischer TW, Kleszczyński K, Hardkop LH, Kruse N, Zillikens D (2013) Melatonin enhances antioxidative enzyme gene expression (CAT, GPx, SOD), prevents their UVR-induced depletion, and protects against the formation of DNA damage (8hydroxy-20-deoxyguanosine) in ex vivo human skin. J. Pineal Res. 54: 303-312. DOI: 10.1111/jpi.12018.
- 181.Ghobadi A, Shirazi A, Najafi M, Kahkesh MH, Rezapoor S (2017) Melatonin ameliorates radiation-induced oxidative stress at targeted and nontargeted lung tissue. J. Med. Phys. 42: 241-244. DOI: 10.4103/jmp.JMP_60_17
- 182.Sliwinski T, Rozej W, Morawiec-Bajda A, Morawiec Z, Reiter R, Blasiak J (2007) Protective action of melatonin against oxidative DNA damage- Chemical inactivation versus base-excision repair. Mutat. Res. Genet. Toxicol. Environ. Mutagen. 634: 220-227. DOI: 10.1016/j.mrgentox.2007.07.013.
- 183.Aydemir S, Akgün SG, Beceren A, Yüksel M, Kumas M, Erdoğan N, Sardaş S, Omurtağ GZ. (2017) Melatonin ameliorates oxidative DNA damage and protects against formaldehyde-induced oxidative stress in rats. Int. J. Clin. Exp. Med. 10: 6250-6261.
- 184. Wu HJ, Liu C, Duan WX, Xu SC, He MD, Chen CH, Wang Y, Zhou Z, Yu ZP, Zhang L, Chen Y (2013) Melatonin ameliorates bisphenol A-induced DNA damage in the germ cells of adult male rats. Mutat. Res. Genet. Toxicol. Environ. Mutagen. 752: 57-67. DOI: 10.1016/j.mrgentox.2013.01.005.
- 185.Erenberk U, Dundaroz R, Gok O, Uysal O, Agus S, Yuksel A, Yilmaz B, Kilic U (2014) Melatonin attenuates phenytoin sodium-induced DNA damage. Drug Chem. Toxicol. 37: 233–239. DOI: 10.3109/01480545.2013.838777.
- 186.Reiter RJ, Sharma R, Ma Q, Rosales-Corral S, Acuna-Castroviejo D, Escames G (2019) Inhibition of mitochondrial pyruvate dehydrogenase kinase: a proposed mechanism by which melatonin causes cancer cells to overcome cytosolic glycolysis, reduce tumor biomass and reverse insensitivity to chemotherapy. Melatonin Res. 2: 105-119. DOI: https://doi.org/10.32794/mr11250033.
- 187. Huang SH, Cao XJ, Wei W (2008) Melatonin decreases TLR3-mediated inflammatory factor expression via inhibition of NF-kB activation in respiratory syncytial virus-infected RAW264, 7 macrophages. J. Pineal Res. 45: 93-100. DOI: 10.1111/j.1600-079X.2008.00560.x.
- 188. Cho SY, Lee HJ, Jeong SJ, Lee HJ, Kim HS, Chen CY, Lee EO, Kim SH (2011) Sphingosine kinase 1 pathway is involved in melatonin-induced HIF-1a inactivation in hypoxic PC-3 prostate cancer cells. J. Pineal Res. 51: 87–93. DOI: 10.1111/j.1600-079X.2011.00865.x.
- 189. Pizarro JG, Yeste-Velasco M, Esparza JL, Verdaguer E, Pallàs M, Camins A, Folch J (2008) The antiproliferative activity of melatonin in B65 rat dopaminergic neuroblastoma cells is related to the downregulation of cell cycle-related genes. J. Pineal Res. 45: 8-16. DOI: 10.1111/j.1600-079X.2007.00548.x.

- 190.Hill SM, Frasch T, Xiang S, Yuan L, Duplessis T, Mao L (2009) Molecular mechanisms of melatonin anticancer effects. Integr. Cancer Ther. 8: 337–346. DOI: 10.1177/1534735409353332.
- 191.Mortezaee K, Najafi M, Farhood B, Ahmadi A, Potes Y, Shabeeb D, Musa AE (2019) Modulation of apoptosis by melatonin for improving cancer treatment updated efficiency: An review. Life Sci. 228: 228-241. DOI: 10.1016/j.lfs.2019.05.009.
- 192. Ferreira GM, Martinez M, Camargo IC, Domeniconi RF, Martinez FE, Chuffa LG (2014) Melatonin attenuates Her-2, p38 MAPK, p-AKT, and mTOR levels in ovarian carcinoma of ethanol-preferring rats. J. Cancer. 5: 728-735. DOI: 10.7150/jca.10196.
- 193.Sahna E, Parlakpinar H, Ozer MK, Ozturk F, Ozugurlu F, Acet A (2003) Melatonin protects against myocardial doxorubicin toxicity in rats: role of physiological concentrations. J. Pineal Res .35: 257-261. DOI: 10.1034/i.1600-079x.2003.00084.x.
- 194.Bilginoğlu A, Aydın D, Özsoy Ş, Aygün H (2014) Protective effect of melatonin on adriamycin-induced cardiotoxicity in rats. Turk. Kardivol. Dern. Ars. 42: 265-273. DOI: 10.5543/tkda.2014.36089.
- 195.Guven A, Yavuz O, Cam M, Ercan F, Bukan N, Comunoglu C (2007) Melatonin protects against epirubicin-induced cardiotoxicity. Acta Histochem. 109: 52-60. DOI: 10.1016/j.acthis.2006.09.007.
- 196.Ghani EA, Kerr I, Dada R (2014) Grade 3 trastuzumab-induced neutropenia in breast cancer patient. J. Oncol. Pharm. Pract. 20: 154-157. DOI: 10.1177/1078155213487394.
- 197.Ozturk M, Ozler M, Kurt YG, Ozturk B, Uysal B, Ersoz N, Yasar M, Demirbas S, Kurt B, Acikel C, Oztas Y (2011) Efficacy of melatonin, mercaptoethylguanidine and 1400W in doxorubicin-and trastuzumab-induced cardiotoxicity. J. Pineal Res. 50: 89-96. DOI: 10.1111/j.1600-079X.2010.00818.x.



This work is licensed under a Creative Commons Attribution 4.0 International License

Please cite this paper as:

Sarkar, S., Chattopadhyay, A. and Bandyopadhyay, D. 2021. Multiple strategies of melatonin protecting against cardiovascular injury related to inflammation: A comprehensive overview. Melatonin Research. 4, 1 (Jan. 2021), 1-29. DOI:https://doi.org/https://doi.org/10.32794/mr11250080.